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TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

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INTERNATIONAL APPLICATION NO.
PCT/CA99/01142INTERNATIONAL FILING DATE
30/11/99 (30 November 1999)PRIORITY DATE CLAIMED
02/12/98 (2 December 1998)

TITLE OF INVENTION METHOD AND APPARATUS FOR MULTIPLE STAGES OF MASS SPECTROMETRY

APPLICANT(S) FOR DO/EO/US Jennifer Campbell, Bruce Collings, Donald J. Douglas

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

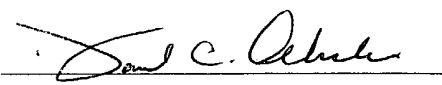
1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to promptly begin national examination procedures (35 U.S.C. 371(f)).
4. ☒ The US has been elected by the expiration of 19 months from the priority date (PCT Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☒ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). **NOTE: WE WILL SUBMIT THE DECLARATION OF 1ST INVENTOR WHEN WE RECEIVE IT.**
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 16 below concern document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A FIRST preliminary amendment.
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:

Amended Pages of Specification
Application Data Sheet
A return postcard

099377 011 0403002
JC18 Rec'd PCT/PTO 01 JUN 2001

U.S. APPLICATION NO. (If known) 097/857234		INTERNATIONAL APPLICATION NO. PCT/CA99/01142		ATTORNEY'S DOCKET NUMBER 021340-00008	
<p>17. <input checked="" type="checkbox"/> The following fees are submitted:</p> <p>BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):</p> <p>Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1000.00</p> <p>International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00</p> <p>International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00</p> <p>International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00</p> <p>International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00</p> <p>ENTER APPROPRIATE BASIC FEE AMOUNT =</p>				CALCULATIONS PTO USE ONLY	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	28 - 20 =	8	X \$18.00	\$	\$144.00
Independent claims	2 - 3 =	0	X \$80.00	\$	\$0.00
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$270.00	\$	\$270.00
TOTAL OF ABOVE CALCULATIONS =				\$	\$1,274.00
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$	
SUBTOTAL =				\$	\$1,274.00
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				\$	
TOTAL NATIONAL FEE =				\$	\$1,274.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property				\$	
TOTAL FEES ENCLOSED =				\$	\$1,274.00
				Amount to be refunded:	\$
				charged:	\$
<p>a. <input type="checkbox"/> A check in the amount of \$_____ to cover the above fees is enclosed.</p> <p>b. <input checked="" type="checkbox"/> Please charge my Deposit Account No. <u>02-2556</u> in the amount of \$ <u>\$1,274.00</u> to cover the above fees. A duplicate copy of this sheet is enclosed.</p> <p>c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>02-2556</u>. A duplicate copy of this sheet is enclosed.</p>					
<p>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.</p>					
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Initial Information Data Sheet

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Application Information

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Title Line Two:: STAGES OF MASS SPECTROMETRY
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Priority Claimed:: Yes

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**Title: METHOD AND APPARATUS FOR MULTIPLE STAGES OF MASS
SPECTROMETRY**

FIELD OF THE INVENTION

This invention relates to multiple stage mass spectrometers
5 which have two mass analyzers, and this invention is more particularly
concerned with both a method of and an apparatus for providing multiple
stages of mass spectrometry (MS^n) capabilities in such spectrometers.

BACKGROUND OF THE INVENTION

Tandem mass spectrometry is widely used for trace analysis
10 and for the determination of the structures of ions. In tandem mass
spectrometry a first mass analyzer selects ions of one particular mass to charge
ratio (or range of mass to charge ratios) from ions supplied by an ion source,
the ions are fragmented and a second mass analyzer records the mass
spectrum of the fragment ions. In a triple quadrupole mass spectrometer
15 system, this effects MS/MS. For example, ions produced in an atmospheric
pressure source, pass through a region of dry nitrogen and then pass through
a small orifice, into a region at a pressure of about 5 torr (0.7 kPa). Ions then
pass through a quadrupole ion guide, operated at a pressure of about 7×10^{-3}
torr (9.1×10^{-4} kPa) into a first quadrupole mass filter, operated at a pressure of
20 about 2×10^{-5} torr (2.6×10^{-6} kPa). Precursor ions mass selected in the first
quadrupole are injected into a collision cell filled with gas, such as argon, to a
pressure of 10^{-4} to 10^{-2} torr (1.3×10^{-5} to 1.3×10^{-3} kPa). The collision cell contains
a second quadrupole ion guide, to confine ions to the axis. Ions gain internal
energy through collisions with the gas and then fragment. The fragment ions
25 and any undissociated precursor ions then pass into a second mass analyzer,
and then to a detector, where the mass spectrum is recorded.

Triple quadrupole systems are widely used for tandem mass
spectrometry. One limitation is that recording a fragment mass spectrum can
be time consuming because the second mass analyzer must step through many
30 masses to record a complete spectrum. To overcome these

limitations, QqTOF systems have been developed. This system is similar to the triple quadrupole system but the second mass analyzer is replaced by a time-of-flight mass analyzer, TOF. The advantage of the TOF is that it can record 10^4 or more complete mass spectra in one second. Thus for applications where a complete mass spectrum of fragment ions is desired the duty cycle is greatly improved with a TOF mass analyzer and spectra can be acquired more quickly. Alternatively for a given measurement time, spectra can be acquired on a smaller amount of sample.

A further known technique is the coupling of electrospray ionization (ESI) to time-of-flight mass spectrometers (TOFMS), and this is an attractive technique for mass spectrometry. ESI is a soft ionization technique capable of forming ions from a broad range of biomolecules, while TOFMS has the well known advantages of rapid mass scanning, high sensitivity, and a theoretically limitless mass range. However, ESI and TOFMS are, in one way, incompatible as a source/analyzer pair: ESI creates a continuous stream of ions and TOFMS requires pulsed operation. Thus in the simplest coupling of ESI to TOFMS there is a very poor duty cycle, with less than 1% of the ions formed being detected and early work in this field was predominantly concerned with increasing the duty cycle.

Within the past two years, literature on ESI-TOFMS has begun to focus on tandem mass spectrometry (MS/MS) with hybrid instruments. The fragmentation of ions in these systems is achieved via traditional methods for collision induced dissociation (CID). Tandem-in-space systems termed quadrupole-TOF's or "Qq-TOF's", as noted above, are analogous to triple quadrupole mass spectrometers - the precursor ion is selected in a quadrupole mass filter, dissociated in a radiofrequency- (RF-) only multipole collision cell, and the resultant fragments are analyzed in a TOFMS. Tandem-in-time systems use a 3-D ion trap mass spectrometer (ITMS) for selecting and fragmenting the precursor ion, but pulse the fragment ions out of the trap and into a TOFMS for mass analysis.

It is sometimes desirable to perform multiple stages of tandem spectrometry termed MS^n . In MS^3 for example, a precursor ion is

selected in a first mass analyzer and dissociated to produce fragment ions. A fragment ion of a particular mass to charge ratio is then isolated and dissociated again to produce fragments of the fragment. The mass spectrum of these is then recorded. Multiple stages of MS are useful when insufficient
 5 dissociation can be produced in a first stage of MS/MS or to elucidate dissociation pathways of complex ions. The latter for example is especially useful to sequence peptides and other biomolecules by mass spectrometry.

The triple quadrupole system and QqTOF system described above provide only one stage of MS/MS and do not allow MS^n . In particular
 10 such systems do not provide for trapping of ions.

In one known proposal, in PCT application WO 98/06481 from Analytica of Brantford, there is a described system including ion trapping. Ions from a source are injected into a multipole ion guide, and ions of one m/z or range of m/z are then isolated in the ion guide by applying resonant
 15 excitation or AC/DC voltages to the ion guide and trapping voltages at either end. The ion is then fragmented in the same ion guide, which can be operated as a linear ion trap (LIT). No mass analyzer is placed before the ion guide. This is a distinct disadvantage, since a multipole ion guide used both for ion isolation and mass analysis has a relatively low resolution. For example, the
 20 present inventors have found that using a LIT as described by Analytica the resolution in isolating an ion is *ca.* 100. With a separate quadrupole mass filter or other mass analyzer before the ion trap the resolution can be many thousand. The relatively low resolution for ions introduced into the multipole ion trap may derive from at least two sources: (1) the pressure is relatively
 25 high (10^{-3} - 10^{-1} torr (1.3×10^{-4} to 1.3×10^{-2} kPa) as described in the PCT application); and (2) in the system described in the PCT application the gas is either nitrogen or air that flows in from the ion source. This has a greater damping effect on ion motion in the LIT than lighter gases such as helium, and gives relatively poor resolution for resonant excitation of ions. Such a system
 30 does not readily enable the pressure and type of gas in the LIT to be adjusted to provide optimum conditions for MS^n .

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Additionally, there have been some recent examples of proposals using resonant excitation in RF-only quadrupoles for CID with fragment mass analysis by TOFMS. Dodonov et al (Rap. Comm. Mass Spectrometry 11, 1649-1656 (1997)) introduced a molecular ion reactor (MIR) consisting of a segmented RF-only quadrupole with a longitudinal electrical field which is operated at a high pressure. Depending on the mode of operation, CID was accomplished through either increasing the RF or DC voltages along the segments. However, no trapping of ions was demonstrated.

Loboda et al (proceedings of the 46th ASMS Conference on Mass Spectrometry and Allied Topics, Orlando, Florida, May 31-June 4, 1998, MOD. 11: 55) modified the RF drive of the collision cell in a Q-TOFMS to apply quadrupolar excitation to ions flowing through the cell, inducing fragmentation. No trapping of ions was demonstrated. It was suggested that a 2D trap might be formed to isolate precursor ions, but it was not stated if this was to be done before or after a stage of mass analysis.

B.A. Thomson et al, in PCT application PCT/CA96/00541, describes a method and apparatus for speeding up the passage of ions through various stages of a mass spectrometer, such as the ion guide and the collision cell. The increase in ion speed is achieved via an axial DC field which can be created through various multipole rod configurations. The axial DC field also aids in the dissociation of ions in collision cells by oscillating the ions axially about their equilibrium positions. However, Thomson states that there is no need to operate at the resonant frequency of the ions or even at a harmonic of the resonant frequency of the ions.

J.D. Watson et al, in an article entitled "*A Technique for Mass Selective Ion Rejection in a Quadrupole Reaction Chamber*" (International Journal of Mass Spectrometry and Ion Processes, 93, 225-235, 1989) described trapping and resonant ejection of an ion from the quadrupole collision cell of a triple quadrupole mass spectrometer. The intent was to study reaction kinetics of trapped ions. While there is no specific teaching of resonant excitation of trapped ions without ejection, there is speculation that this might be possible.

In mass spectrometry, the linear ion trap has remained relatively unexplored. U.S. patents 4,755,670 and 5,420,425, both assigned to the Finnigan Corporation, relate to a Fourier transform quadrupole and new ion trap

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geometries respectively, and they both mention a LIT. U.S. patent 5,179,278 (D.J. Douglas) suggests using a LIT as an "ion bottle" to improve the duty cycle of a 3D ITMS.

5 SUMMARY OF THE INVENTION

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In accordance with a first aspect of the present invention, there is provided a method of analyzing a stream of ions, the method comprising:

- 5 (1) subjecting a stream of ions to a first mass analysis step, to select ions having a mass-to-charge ratio in a first desired range;
- (2) passing ions in the selected range into a radio frequency linear ion trap containing gas;
- (3) trapping the selected ions in the linear ion trap and exciting the ions to cause collisions with the ambient gas and fragmentation;
- 10 (4) subjecting the fragmented ions to a secondary excitation, different from the first excitation, to cause excitation and fragmentation of selected fragment ions; and
- (5) passing the ions out of the linear ion trap and subjecting the ions to a further mass analysis step to determine the mass spectrum of the ions.
- 15

Passing the ions, in step (2) into the radio frequency ion trap can be done either: with a relatively low energy, so no fragmentation occurs in the LIT until additional excitation is applied; or with a relatively high energy in the axial direction, so that fragmentation occurs simply due to the high energy of the ions entering the LIT and colliding with the gas.

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Thus a variant of the basic method of the present invention comprises passing the ions into the linear ion trap with sufficient energy to promote collision induced dissociation, said energy providing the excitation of (3), whereby step (3) comprises applying a signal to the linear ion trap to trap ions, before subjecting the ions to the further mass analysis of step (5).

25

In either case, once the ions have entered the LIT, have been excited, by either technique, to cause fragmentation, one then has fragmentions, with any remaining precursor ions trapped in the LIT. These ions can then be discharged for further mass analysis, or subject to multiple steps of mass selection and excitation to cause fragmentation, before being discharged for the final mass analysis step.

30

Thus, the method advantageously includes, in step (4), subjecting the fragmented ions to a secondary excitation, different from the first excitation, to cause excitation and fragmentation of selected fragment ions

(MS³). This can be repeated to achieve further steps of MSⁿ (n greater than 3). Further, prior to the additional step of secondary excitation, applying a signal to the linear ion trap, to select ions having a mass-to-charge ratio in a second desired range, wherein the secondary excitation step comprises exciting ions in
5 the second desired range.

Thus, the method can include, while trapping the ions in the linear ion trap, effecting multiple cycles of:

- (1) selecting ions having a mass-to-charge ratio in a desired range; and
- 10 (2) exciting the selected ions to cause fragmentation.

The ions can be excited in the linear ion trap by providing an additional signal to the linear ion trap.

The further mass analysis step of step (5) can be carried out either in a quadrupole mass analyzer, or in a time of flight mass analyzer. For a
15 time of flight mass analyzer, this can be arranged with its axis perpendicular to the axis of the linear ion trap.

Preferably, the first mass analysis step is carried out in a quadrupole mass analyzer which is coaxial with the linear ion trap.

More preferably, the method includes, prior to exciting the
20 ions in step (3), subjecting the trapped ions to a signal comprising a plurality of excitation signals uniformly spaced in the frequency domain and having a notch, wherein the notch covers a desired frequency band and there are no excitation signals in the frequency band of the notch, and wherein the excitation signals have sufficient magnitude to excite and eject ions except for
25 ions having an excitation frequency falling within the frequency band of the notch. For the case where the frequency of the trapping RF signal is 1.0 MHz, this can be achieved by applying a combination of signals having sine waves with frequencies in the range 10 to 500 kHz and spaced at 500 Hz intervals, and the frequency band of the notch then has a width of typically 1-10 kHz and is
30 centered on the resonant frequency of an ion of interest. More generally, where the trapping RF frequency is f , then the auxiliary frequencies should be up to $f/2$.

The linear ion trap can have a pair of opposed x rods and a pair of opposed y rods, and then a main RF drive is connected to the x and y

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rods of the linear ion trap and an auxiliary drive is connected to at least one pair of rods of the linear ion trap. For example, the auxiliary drive is connected between the x and the y rods of the linear ion trap through a transformer, and the main RF drive is connected directly to the x rods of the linear ion trap and, through a coil of the transformer to the y rods. Alternatively, the auxiliary drive can be connected between the x rods. The apparatus preferably then includes an arbitrary waveform generator connected to the auxiliary drive, for applying a selected waveform to the linear ion trap to excite ions therein.

10 DESCRIPTION OF THE DRAWINGS

For a better understanding of the present invention and to show more clearly how it may be carried into effect, reference will now be made, by way of example, to the accompanying drawings in which:

Figure 1 is a schematic diagram of a mass spectrometer apparatus in accordance with the first embodiment of the present invention;

Figure 2a is a schematic diagram of a mass spectrometer including a TOF in accordance with the second embodiment of the present invention;

Figure 2b is a schematic diagram of a mass spectrometer including a TOF, according to a third embodiment of the present invention and similar to Figure 2a, but without a first mass resolving quadrupole;

Figure 3 is a schematic diagram showing coupling of an auxiliary drive to quadrupole rods;

Figure 4 is a diagram showing variation of voltages in various elements of the spectrometer of Figure 2 over a cycle;

Figures 5a and 5b show spectra from a solution of renin substrate, showing isolation of a selected charge state;

Figure 6 is an isometric 3-dimensional view showing variation of ion intensity with channel number and excitation frequency;

Figures 7a and 7b are graphs showing variation of intensity

against frequency.

Figures 7c and 7d are graphs showing variation of intensity against frequency for different pressures in the chamber of Figure 2b;

Figure 7e is a graph showing variation of resolution with
5 pressure for different excitation voltages for the apparatus of Figure 2b;

Figure 8 is an isometric 3-dimensional view showing variation of the intensity of reserpine precursor ion with the auxiliary voltage;

Figures 9a and 9b are graphs showing similar plots to Figure 8 with the auxiliary voltage at the resonant frequency and 2-5 kHz below resonant frequency;

Figure 10 shows a variation of precursor and fragment intensity with excitation period; and

Figure 11 is a series of graphs demonstrating MS³ in the apparatus of Figure 2b.

15 DESCRIPTION OF THE PREFERRED EMBODIMENTS

Referring first to Figure 1, a mass spectrometer is indicated generally by the reference 10. Ions are generated by an ion source 12, which is a pneumatically assisted electrospray, and pass through a dry nitrogen "curtain gas", indicated at 14. The ions then pass through an orifice in plate 16, and then through a further orifice in a skimmer 18, into a first quadrupole rod set Q0.

The rod set Q0 is located in a first chamber 22 which is connected to a turbo molecular pump, with the connection indicated at 24. Although not shown, in known manner, the turbo molecular pump 24 is backed up by a rotary vane pump, which can also be connected to the region 25 between the orifice plate 16 and the skimmer plate 18. Alternatively the region between the orifice and skimmer plates 16, 18 can be evacuated by a separate rotary vane pump.

The turbo molecular pump 24 maintains a pressure of 7×10^{-3} torr (9.1×10^{-4} kPa) in the chamber 22, while a pressure of 2 torr (0.3 kPa) is maintained between

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the orifice and skimmer plates 16, 18. The rod set Q0 has just an RF voltage applied to it, so that it operates as an ion guide.

Ions then pass through into a main chamber 26 of the mass spectrometer. Within the main chamber 26, there are located first, second and
5 third quadrupole rod sets, indicated at Q1, Q2 and Q3. A detector 36 is provided at the exit from the final rod set at Q3.

As indicated at 30, a connection to a suitable turbo molecular pump would be provided, again backed by the same rotary vane pump that backs turbo molecular pump 24. The pump 30 maintains a pressure of 2×10^{-5}
10 torr (2.6×10^{-6} kPa) in the main chamber 26.

The central quadrupole rod set Q2 is enclosed in a chamber or housing 28 and is provided with a connection for a gas (not shown), so that a higher pressure can be maintained typically at around 1-7 millitorr (1.3×10^{-4} to 9.1×10^{-4} kPa).

Now, in accordance with the present invention, the housing or enclosure 28 with the rod set Q2 forms a linear ion trap. For this purpose, conductive plates with apertures are provided at the ends of the housing 28, which may be either separate from the housing 28 or integral therewith. These comprise an entrance plate 32 and an exit plate 33. The plates 32, 33 are
15 20 conductive, insulated from another and connected to voltage sources 34.

Downstream from the housing 28 is a third quadrupole rod set, Q3, configured as a mass analyzer. For operation as a conventional triple quadrupole MS/MS system, the quadrupole rod sets Q0, Q1, Q2 and Q3 would be connected to conventional voltage sources, for supplying DC and RF
25 voltages as required.

In use, ions generated from the ion source 12 pass into the quadrupole ion guide Q0. As noted, this is supplied with just RF voltages, to operate as an ion guide. Ions then pass through Q0 into the first quadrupole rod set Q1. This is supplied with suitable RF and DC voltages to operate as a
30 mass filter, to select ions with a desired m/z ratio.

A mass selected precursor ion from the first rod set Q1 is then injected into the collision cell 28, to produce fragment ions as is

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known, by collision with a gas in the collision cell. If the energy with which the precursor ions enter the collision cell is low, they remain largely undissociated. The extent of ion fragmentation can be controlled by changing the injection ion energy and by changing the type and the pressure
5 of the gas in Q2.

However, a blocking potential is applied to the exit plate 33. Consequently, these fragment ions are not immediately transmitted to the downstream rod set Q3. A blocking potential is then applied to the inlet 32 of the collision cell 28, to prevent additional ions entering the collision cell
10 28.

Under these conditions, the collision cell 28 forms a radio frequency linear ion trap (LIT). The precursor ion or the fragment ion of a particular mass to charge ratio (m/z) can then be isolated in the collision cell or LIT 28 by a number of methods, such as resonant ejection of all other
15 ions, application of RF and DC voltages to the LIT to isolate an ion at the tip of a stability region, or ejection of ions with an m/z lower than that of the selected ion by increasing the RF voltage or other known means.

After isolation the selected ion can then be excited by resonant excitation or other means to produce fragments of the selected,
20 fragment ions; thus the original ions from source 12 are dissociated to produce fragment ions, and a selected fragment ion can be further fragmented to produce fragments of fragment ions.

The blocking potential at the exit 33 of the collision cell 28 can then be lowered to transfer the ions to the third quadrupole Q3. When
25 ions are being transferred to the third quadrupole, a stopping potential is applied to the entrance plate 32. Quadrupole Q3 is operated, with suitable RF and DC voltages, to record a spectrum at the detector 36. It will be appreciated that the trapping isolation and fragmentation cycle can be repeated more than once, to provide MS^n capabilities.

30 Reference will now be made to Figure 2a, which shows an apparatus similar to Figure 1 but with the third quadrupole Q3 replaced by a time of flight instrument, indicated at 40. Otherwise, for simplicity and

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brevity, like components in Figure 2a are given the same reference numeral as in Figure 1, and description of these components is not repeated.

In Figure 2a, the time of flight device 40 is connected to the exit plate 33 of the collision cell 28. In known manner, the time of flight device 40 includes a connection 42 to a pump for maintaining a vacuum at 5×10^{-7} torr (6.5×10^{-8} kPa). It includes a repeller grid 44 and other grids indicated schematically at 46, for collecting ions entering the TOF 40 and transmitting a pulse of ions. The TOF device 40 here is a reflectron and includes grids 48 for reflecting the ion beam, which is then detected by a detector 49. A linear TOF may also be used, as shown in Figure 2b.

The apparatus in Figure 2a would be operated in an essentially similar manner to that of Figure 1. The principal difference is that the TOF can record 10^4 or more complete mass spectra in one second. Thus for applications where a complete mass spectrum of fragment ions is desired the duty cycle is greatly improved with a TOF mass analyzer 40 and spectra can be acquired more quickly. Alternatively, for a given measurement time, spectra can be acquired on a smaller amount of sample.

While three-dimensional (3-D) traps (IT) have been provided in spectrometers including a TOF final stage, a two-dimensional (2-D) trap has several advantages over the 3-D trap. Firstly, because there is no quadrupolar electric field in the z direction, the ion injection and extraction efficiencies can be nearly 100%. As fewer ions are lost in the processes of filling and emptying the trap the sensitivity of the Linear Ion Trap Time Of Flight Mass Spectrometer (LIT/TOFMS) can be greater than that of the IT/TOFMS (an ESI source, a 3-D ion trap mass spectrometer and a TOFMS).

Because of the increased trapping volume of the LIT, a greater number of ions (N_{2-d}) can be trapped in a LIT than in a 3-D trap (N_{3-d}). The increase in ion capacity is given by

$$\frac{N_{2-d}}{N_{3-d}} = \frac{r_0^2 l}{z_0^3} \quad (1)$$

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where l is the length of the LIT, r_0 is the field radius of the LIT, and z_0 is the z direction field radius of the 3-D ion trap mass spectrometer. For example, given that, in the apparatus described below, l is 20 cm, r_0 is 0.4 cm, and a typical z_0 for a commercial trap is 0.707 cm, the linear ion trap of the present invention has almost an order of magnitude increase in ion capacity. The higher ion capacity increases the concentration linear dynamic range of the LIT/TOFMS relative to the IT/TOFMS.

In addition to providing a greater trapping volume, the LIT can be operated in all of the modes for mass isolation and MS/MS of a 3-D ITMS. Ion motion in the RF quadrupole fields of both the quadrupole rod set and the quadrupole ITMS geometry are identical and described mathematically by the solutions to the Mathieu equation. Ion motion is decoupled in each coordinate, u , of the quadrupole field - x and y in the RF-only quadrupole and the x - y plane and z in the 3-D ITMS. Parameters for which motion is stable in each coordinate are determined by the Mathieu parameter, q_u ,

$$q_u = \frac{4V_{rf}}{\frac{m}{z} \Omega^2 u_0^2} \quad (2)$$

where V_{rf} is the applied RF voltage from an electrode to ground (0 to peak), $\frac{m}{z}$ is the mass-to-charge ratio of the ion, u_0 is the field radius of the device for that co-ordinate, and Ω is the angular frequency of the trapping RF drive. In the commonly used first stability region, ions for which $0 \leq q_u \leq 0.9$ have stable trajectories in the quadrupole device. Thus, if V_{rf} and Ω are fixed there is a lower limit to the $\frac{m}{z}$ of ions which have stable trajectories in the trap.

The frequencies of ion motion are given by

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$$\omega_n = (2n + \beta) \frac{\Omega}{2} \quad (3)$$

where n is an integer, $-\infty < n < \infty$, and β is a function of q_u . If $\beta \leq 0.4$, ($q_{x,y} \leq 0.6$ in an RF only quadrupole) then the adiabatic approximation is valid and ion motion in the quadrupole field is like that of a charged particle moving
 5 in a harmonic "pseudopotential" well of depth

$$D_u = \frac{q_u}{8} V_{rf} \quad (4)$$

Each $\frac{m}{z}$ thus has a unique fundamental resonant frequency ω_0 ($n = 0$ in equation (3)), given approximately by

$$\omega_0 = \frac{q_u}{\sqrt{8}} \Omega \quad (5)$$

- 10 The motion of the ion in the LIT can be excited through the application of an auxiliary voltage on one set of pole pairs in the x or y direction. This oscillating voltage, V_a , has the form

$$V_a = A_a \sin \omega_a t \quad (6)$$

- where A_a and ω_a are the amplitude and frequency of the auxiliary voltage,
 15 and t time. Application of the auxiliary voltage at the resonant frequency of an ion causes the amplitude of its oscillation to increase linearly with time. If the amplitude exceeds r_0 (or equivalently, energy increase from resonant absorption is greater than D_u) the ion will be ejected from the trap. In the presence of a background neutral gas, the excited ion motion will result in
 20 an increase in the number and energy of collisions. As kinetic energy is transferred to ion internal energy, the ion may reach its critical energy for collision induced dissociation (CID) and fragment.

Reference will now be made to Figure 2b, which shows an alternative embodiment. This was designed without the initial, mass resolving quadrupole Q1, to provide experimental data on the performance of the LIT. It also includes a linear TOF section, to provide LIT/TOFMS.

5 The LIT/TOFMS was designed to be flexible with three modes of operation: (i) continuous flow-TOFMS, in which the products of ESI can be analyzed without trapping or fragmentation; (ii) trap-TOFMS, in which the combination of trapping and pulsing ions can be used to enhance instrumental duty cycle; and (iii) MS/MS-TOFMS in which the fragmentation spectra for
10 isolated precursor ions are recorded via TOFMS. Switching between modes is a simple matter of changing the parameters which control timing, trap entrance and exit potentials, and excitation frequencies and amplitudes.

In Figure 2b, the spectrometer is indicated generally at 50. Ions are generated by pneumatically assisted electrospray at 52 and pass
15 through a dry nitrogen curtain gas 54, a 0.25 mm diameter sampling orifice in an orifice plate 56, a 0.75 mm diameter orifice in the skimmer 58, and into a first RF-only quadrupole Q0. The region between the skimmer and the orifice is evacuated by a rotary vane pump as indicated at 62, to a pressure of 2 torr (0.3 kPa). A second quadrupole rod set is indicated at Q2. For consistency with
20 Figure 2a, the designation Q2 is used, although there is no Q1 in Figure 2b.

The RF-only quadrupoles Q0, Q2 are separated by a 1mm diameter interquad aperture 64 (IQ). The first quadrupole, Q0, is 5 cm long and the second Q2, which acts as the LIT, is 20 cm long. Both quadrupoles have field radii of 4.0 mm and are operated by the same main RF drive, which has a
25 maximum V_{rf} of 5000 V and a drive frequency, $\Omega = 2\pi f$ where f is 1 MHz. There are 12.5 pF capacitors between the RF drive supply and the rods of Q0; thus the RF voltage on Q0 is ca. one half that of Q2. The DC offsets of the quadrupoles are individually set and for all the experiments here these are Q0=10 V and Q2=0 V. The pressure in the LIT can be varied from 1.5 to 7.0
30 mTorr (2×10^{-4} to 9.1×10^{-4} kPa) by adding gas. The region surrounding the LIT

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provided by Q2 is connected to a turbomolecular pump, as indicated at 66. The LIT chamber is indicated at 68.

5 A TOF chamber 70 is coupled orthogonally to the LIT chamber 68 via four lenses, L1-L4. L1 (aperture diameter 0.75 mm) serves as the exit of the LIT chamber 68 and the differential pumping aperture between the LIT and the TOF chambers. The three lenses, L2, L3 and L4, have apertures of 2 mm diameter and are used to focus the ion beam into the source region of a two stage, 1 m long, TOFMS. The TOF chamber 70 is held at a pressure of 1.2×10^{-6} torr (1.6×10^{-7} kPa) or less by a turbomolecular pump. Separate rotary
10 vane pumps are used to pump the region between the orifice and skimmer and to back the turbo pumps.

In the TOF source region, in known manner there are a repeller grid 72, a middle TOF grid 74 and a final TOF grid 76. The ion source was operated near ground potential and the flight tube was floated at a
15 negative high potential, typically 2.0kV. To reduce distorting effects of the floating voltage a shielding grid 78 was placed 4.2 mm behind the middle TOF grid 74. An additional shielding grid 80 was placed around the repeller grid 72 and the middle TOF grid to reduce the effects of stray fields on ions entering the source region. Ions are accelerated in the TOF in a direction orthogonal to
20 that of the quadrupoles. Thus, the system is termed an orthogonal acceleration TOF (oa-TOF).

There is no potential difference between the Q2 offset and the source region, thus the axial ion energy in the TOF source is < 1 eV. The repeller grid 72 of the TOF is pulsed from an offset of 0 V to an amplitude of \approx
25 200-300 V using a high voltage (HV) pulser (rise time < 18 ns). The amplitude of the HV pulse is adjusted to achieve maximum resolution for the ion acceleration energy. Because the ions enter the source region midway between the repeller grid 72 and grid 74, the acceleration energy is given by one half of the amplitude of the HV pulse minus the negative float potential. The
30 experimental HV pulse amplitudes that gave the best resolution were found to equal those calculated to give space focussing for the set acceleration energies. The HV pulse width is set to be greater than

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the time for the ions with the highest $\frac{m}{z}$ to exit the TOF acceleration region. This width is much less than the flight time which defines the TOFMS scanning rate, typically 10 μ s and 100 μ s respectively. After the HV pulse is complete and during the ion flight time, the repeller plate 72 voltage is set to a potential which allows for ion transmission into the source region. The duty cycle of the oa-TOFMS is thus given by the ratio of the source filling time to the time between the pulses to the repeller plate 72. Because this duty cycle is increased if the ions move more slowly through the source region it is preferable that the coupling of the LIT to the TOFMS incorporate a method to ensure low energy ions enter the source.

Duty cycle, resolution, and sensitivity are all increased through the combination of the orthogonal acceleration coupling geometry with collisional cooling in RF-only quadrupoles operated at relatively high pressures. In the z direction of the quadrupole, dampening of translational energy creates a slower, higher ion density beam. In addition to increasing the duty cycle, a slower beam gives a higher ion density to each pulse accelerated into the flight tube, thus enhancing sensitivity. Energy dampening in the x, y direction also occurs, causing the ions to move to the center of the quadrupole rods. The resultant beam has a small spatial and energy spread in the radial direction, which improves resolution in the TOFMS.

For these benefits from collisional cooling to be realized, it is necessary to minimize any defocusing of the beam as it travels from the LIT to the source region of the TOFMS. In particular, any stray fields from the float potential which penetrate into the source region will cause broadening or deflection of the ion beam, thereby degrading resolution. For this reason, the various shielding grids 78, 80 are provided.

For the study of biomolecules, which often have large collision cross sections, the flight tube must have a pressure which is low enough for the mean free paths (λ) of the ions to be longer than the flight tube. Otherwise collisions between ions and residual gas result in a

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substantial loss in resolution in the TOFMS. Nitrogen was added to the flight tube to increase the pressure over the range 1.2×10^{-6} torr (1.6×10^{-7} kPa) to 5×10^{-5} torr (6.5×10^{-6} kPa), corresponding to a decrease in the mean free path for the +13 charge state of cytochrome *c* (collision cross section $\sim 1700 \text{ \AA}^2$) from ~ 106 cm to ~ 4 cm. The resolution in the TOFMS spectrum degraded from 600 to 30 with a ca. 25 x increase in the number of collisions. At 1.2×10^{-6} torr (1.6×10^{-7} kPa), the probability of the ion undergoing zero collisions in the flight path is 52% and of one collision 34%. Extrapolation of a graph of resolution vs. λ^{-1} shows that at zero pressure the resolution is improved ca. 6% over that of 1.2×10^{-6} torr (1.6×10^{-7} kPa).

A schematic of the RF operation for the LIT is shown in Figure 3. The master clock for the LIT/TOFMS is provided by a two channel arbitrary waveform generator 82 (AWG). Each channel of the AWG 82 provides a maximum amplitude (0 to peak) of 12 V. The AWG 82 is connected to an auxiliary drive (Aux. Drive) 84, which in turn is connected by a bipolar transformer 85 to the *y* rods. A main RF drive 86, as shown, is connected directly to the *x* rods, with one connection being through the transformer 85 to the *y* rods.

As shown in Figure 4 the complete MS/MS cycle takes 20 ms to complete. It involves changing the potentials on the interquad aperture (IQ) 64 and exit aperture L1, control of the auxiliary driver 84 which connects the output of the AWG 82 to the quadrupole rods Q2, and the TOFMS pulsing (TOF).

The first phase of the cycle is ion injection. A synchronization pulse from the AWG 82 triggers a pulse generator (not shown) which controls the potential on IQ 64, which is maintained at a potential (~ 7 V) indicated at 100 to pass ions for a set injection time (typically 5 ms as shown in Figure 4) and a stopping potential 102 (12 V) for the remaining 15 ms of the scan. In addition this injection time serves as a thermalization period. For the ions that were studied, fragmentation spectra were independent of orifice skimmer potential difference, suggesting that any ion heating in the ion sampling region has equilibrated during the

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For the trapping period both IQ 64 and L1 are held at stopping potentials (12 V) as shown at 102 and 107, with the stopping potential being applied to IQ 64 after the injection period. It has been shown previously and was experimentally verified for this system, that the LIT has a near 100% trapping efficiency for periods of at least up to 200 ms. All data were recorded with trapping times much less than 200 ms so there is no

- 20 -

need to consider trapping losses.

The last phase of the MS/MS cycle is the detection of fragment ions. As shown in Figure 4, L1, which is controlled by channel 1 of the AWG, is held at a stopping potential 107 (+12) for the first 13 ms of the MS/MS scan and at a potential 108 (-10V) to transmit ions for a set trap emptying time, typically 7 ms. In addition, channel one of the AWG 82 gates a pulse generator (not shown) which is used to trigger the TOF HV pulsing and the detection electronics. Thus, only when the trap is being emptied are TOF scans acquired. As indicated at 110, the TOF repeller grid 72 is turned off during the front 13ms of the cycle and during a trap empty period of 7ms is excited at the scanning rate of 10 kHz as indicated at 112. As the TOF scanning rate is typically 10 kHz, there are 70 TOF scans for each empty cycle. The time to fill the source region is typically 10 μ s giving an MS/MS duty cycle determined from separate TOF and quadrupole duty cycles as follows:

15 TOF duty cycle:

$$\frac{\text{number of TOF scans} \times \text{time to fill TOF source region}}{\text{time for whole cycle}} \times 100\% = \frac{70 \times 0.01}{20} = 4\%$$

quadrupole duty cycle:

$$\frac{\text{injection time}}{\text{time for whole cycle}} \times 100 = 25\%$$

overall MS/MS duty cycle:

$$25\% \times 4\% = 1\%$$

The following materials were used in the tests detailed below:

20 Reserpine, renin substrate tetradecapeptide, and cytochrome c (from horse heart) from Sigma (St. Louis MO., U.S.A.). Unless

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otherwise noted, solutions were as follows: reserpine (MW=608), 1 μ M in acetonitrile (HPLC grade, Fisher Scientific, Nepean, Ont. Canada); renin substrate (mw=1759), 5 μ M in 85:10 water:methanol with 5% acetic acid (HPLC grade, Fisher Scientific, Nepean, Ont. Canada); and cytochrome c
5 (mw=12300) 5 μ M in 50:50 water:methanol with 5% acetic acid. The solution flow rate for all experiments was 1 μ L/min. Gases were from Praxair, (Vancouver, B.C., Canada) with manufacturer's stated purities of 99.999% (N_2 , UHP grade) and 99.995% (Kr, UHP grade).

The use of the LIT to enhance the duty cycle of the TOFMS
10 was demonstrated with a storage experiment using ions of cytochrome c. In this experiment, IQ 64 is always set to pass ions. L1 is held at stopping potential for varying lengths of time. The time between the lowering of the potential on L1 and the scanning of the TOFMS is varied to determine the time for the densest portion of the trapped beam to reach the accelerating
15 region. There was a single TOF scan for each trapping period.

On average the delay required between lowering L1 and the TOF scanning was 60 μ s and the TOF accelerating pulse width was 10 μ s. When operated in continuous flowing conditions the TOFMS 50 had an intensity of 2.2 ion counts per pulse. If a stopping potential is applied to L1
20 for the last 40 μ s of the 100 μ s flight time, ion counts per pulse were found to triple to 6.6. In effect, this prevents premature entry and subsequent loss of ions in the source region between grids 72, 74; instead, the ions are trapped in Q2, enabling the total number of ions to build up, leading to an increased number of ions per pulse. It is important to note that this
25 sensitivity enhancement occurs without any sacrifice in TOFMS scanning time. Similarly, if the repetition rate is decreased to 5 kHz that is the time between TOF MS pulses is increased to 200 μ s and the stopping potential applied to L1 for the last 100 μ s of the flight scan, the ion intensity increased to 8.5 ions per pulse, which is almost double the 4.4 ions which would be
30 detected in the same time period were no trapping used. Using trapping to create a denser, low energy beam gives overall higher sensitivity than co-adding with continuous flowing conditions.

25 Figure 6 shows the raw data for an MS/MS experiment
which demonstrates the variation in the recorded spectra of renin substrate
as the frequency of the auxiliary voltage is varied. The spectra are plotted in
channel numbers, where each channel is 20 ns wide and channel 0
represents a flight time of 30 μ s. Figure 6 shows the variation of intensity
30 with both channel # and frequency of auxiliary voltage applied to Q2. In
Figure 6, the m/z of an ion is related to the channel number, n , by the

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following equation:

$$n = a\sqrt{\frac{m}{z}} + b, \text{ where } a \text{ and } b \text{ are constants.}$$

The +3 charge state ($\frac{m}{z} = 587$) was isolated via application of the broadband waveform for 4ms, and the precursor was excited for 4ms (amplitude 1.5 V, $q_y \sim 0.623$, $\omega_o \sim 220$ kHz). As expected, the intensity of the precursor ion shown at 120, falls as the fundamental frequency $\omega_a = \omega_o$ of its motion is approached and rises again for $\omega_a > \omega_o$. Correspondingly, the major fragmentation, shown at 122, peaks at $\omega_a = \omega_o$. This plot is shown in more detail in Figure 7a which plots the intensity of the precursor ion 120 and sum of fragment ions, indicated at 124, versus excitation frequency. The excitation width for this system is 3.0 kHz; thus the mass resolution of the excitation in this experiment is $73 \left(\frac{f}{\Delta f} = \frac{m}{\Delta m} \right)$. As is evident from Figures 6

and 7a, resonant excitation induces a significant degree of fragmentation but without adequate knowledge of the charge states of these fragments of renin substrate +3 it is difficult to comment on the total fragmentation efficiency of the system. Figure 7b plots the precursor and fragment ion intensities at 126 and 128 respectively, as a function of frequency for the excitation of the +1 charge state of reserpine for 4ms (amplitude 2.4 V, $q_y \sim 0.51, \omega_o = 181$ kHz). Fragmentation efficiency can be calculated from the ratio of the sum of the intensities of all fragment ions to the intensity of the precursor ion prior to the application of the excitation; for reserpine (Figure 7b) it is found to be $\sim \frac{3750}{6250} \times 100 = 60\%$. A higher excitation resolution is possible if one is willing to sacrifice fragmentation efficiency and duty cycle through the use of a lower excitation amplitude in conjunction with a longer excitation period.

Due to the relatively high q_y setting used, the calculated low
 25 mass cutoffs are relatively high. For instance for the data plotted in Figure

30 Excitation of an ion in a trap leads to a competition between

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ion ejection and ion fragmentation. For ejection the important parameters are q_y , the mass and charge of the precursor ions, the pressure and mass of the collision gas, and the collision cross section of the precursor ion, as well as the duration and amplitude of the auxiliary voltage. Fragmentation, however, is
 5 also dependent upon the structure of the precursor ion, the effectiveness of the transfer of kinetic energy to internal energy through collisions, and the time scale for the unimolecular dissociation of the precursor ion. The details of this competition have been discussed for 3-D ITMS previously. There follows a discussion on tests to demonstrate how operating parameters for the
 10 apparatus of Figure 2b affects competition between ejection and fragmentation.

Figure 8 shows the effect on singly charged reserpine ions of increasing the amplitude of the auxiliary voltage, as plotted against intensity and channel number. While a threshold voltage is necessary to induce
 15 fragmentation, as the amplitude increases ejection dominates and no fragmentation is observed. The precursor ion is indicated at 130, and fragments at 132, 134. Figure 9a shows a similar plot for the same experiment for the +3 charge state of renin substrate, with the precursor indicated at 136 and the sum of the fragments at 138. Figure 9b demonstrates the effect of increasing the
 20 amplitude of the excitation voltage at $\omega_a = \omega_o - 2.5$ kHz, with the precursor indicated at 140 and the sum of the fragments at 142. The extent of fragmentation increases by 50% and the excitation amplitude for which fragmentation is one half of its maximum value increases by 40% when off resonant excitation is used. Similar increases in the extent of fragmentation and
 25 the amplitude for which fragment ions could be detected were observed for all ions and charge states examined.

The fragmentation spectrum recorded with low amplitude resonant excitation ($A = 1.5$ V, $\omega_a = \omega_o$) was similar to that recorded in a triple quadrupole mass spectrometer system with 15 eV laboratory injection energy
 30 and a 20 cm long collision cell filled with 1 mTorr (1.3×10^{-4} kPa) of Krypton (The triple quadrupole is described in Y-L. Chen, B.A. Collings, and D.J. Douglas,

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Figure 11 demonstrates MS³ in a linear ion trap, and shows a series of spectra, identified as Figures 11a-11e. The data was recorded on the instrument shown in Figure 2b, and the MS³ timing cycle was similar to that shown in Figure 4. The products of the ESI of renin substrate, injected for 5 ms are shown in Figure 11a. Figure 11b demonstrates the isolation of the +3 charge state ($\frac{m}{z} = 587$) in Q2 via application of a broadband notched waveform for 4 ms. In Figure 11c an MS/MS fragmentation pattern, similar to the plot of Figure 6 is shown. Fragmentation was achieved through the application of a small amplitude sinusoidal oscillation for 1 ms. The next step in the MS³ is the isolation of the dominant fragment, the doubly charged fragment with $\frac{m}{z} = 726$. Figure 11d demonstrates the result of a 4 ms broadband notched waveform designed to isolate this dominant fragment,

CLAIMS:

1. A method of analyzing a stream of ions, the method
5 comprising:
 - (1) subjecting a stream of ions to a first mass analysis step, to select ions having a mass-to-charge ratio in a first desired range;
 - (2) passing ions in the selected range into a radio frequency linear ion trap (Q2) containing a gas;
 - 10 (3) trapping the selected ions in the linear ion trap (Q2) and exciting the ions to cause collisions with the gas and fragmentation;
 - (4) subjecting the fragmented ions to a secondary excitation, different from the first excitation, to cause excitation and fragmentation of selected fragment ions; and
 - 15 (5) passing the ions out of the linear ion trap (Q2) and subjecting the ions to a further mass analysis step to determine the mass spectrum of the ions.
2. A method as claimed in Claim 1, which includes, prior to the
20 additional step of secondary excitation, applying a signal to the linear ion trap (Q2), to isolate ions having a mass-to-charge ratio in a second desired range, wherein the secondary excitation step comprises exciting ions in the second desired range.
- 25 3. A method as claimed in Claim 2, which includes, while trapping the ions in the linear ion trap (Q2), effecting multiple cycles of:
 - (1) isolating ions having a mass-to-charge ratio in a further desired range; and
 - (2) exciting the isolated ions in the further desired range
30 to cause fragmentation.
4. A method as claimed in Claim 1, 2, or 3 wherein step (2) comprises passing the ions into the linear ion trap (Q2) with sufficient energy to promote collision induced dissociation, the said energy providing the

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excitation of step (3), whereby step (3) comprises applying a signal to the linear ion trap (Q2) to trap ions before subjecting the ions to the further mass analysis of step 4).

- 5 5. A method as claimed in Claim 1, 2, 3 or 4 which comprises exciting the ions in the linear ion trap (Q2) by providing an additional signal to the linear ion trap (Q2) to effect resonant radial excitation of ions.
- 10 6. A method as claimed in any preceding claim, wherein the further mass analysis step of step (5) is carried out in a quadrupole mass analyzer (Q3).
- 15 7. A method as claimed in any of claims 1 to 6, wherein the further mass analysis step of step (5) is carried out in a time of flight mass analyzer (40).
- 20 8. A method as claimed in Claim 7 wherein the further mass analysis step is carried out in a time of flight mass analyzer (40) arranged with its axis perpendicular to the axis of the linear ion trap (Q2).
- 25 9. A method as claimed in Claim 1 wherein each mass analysis step is carried out in one of: a linear quadrupole (Q3); a linear time of flight analyzer (40); a reflectron time of flight analyzer; a single magnetic sector analyzer; a double focusing two sector mass analyzer having an electric sector and a magnetic sector; a Paul trap; a Wien filter; a Mattauch-Herzog spectrograph; ion cyclotron mass spectrometer; and a Thomson parabolic mass spectrometer.
- 30 10. A method as claimed in Claim 6, 7, 8, or 9, wherein the first mass analysis step is carried out in a quadrupole mass analyzer (Q1) which is coaxial with the linear ion trap (Q2).
11. A method as claimed in Claim 1, which includes, prior to exciting the ions in step (3), subjecting the trapped ions to a signal comprising a

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17. An apparatus (10), for effecting mass analysis and fragmentation of an ion stream, the apparatus comprising:
an input (12) for an ion stream;
a first mass analyzer (Q1);
5 a radio frequency linear ion trap (Q2);
a final mass analyzer (Q3); and,
an auxiliary drive (84) connected to the radio frequency linear ion trap (Q2) for effecting multiple excitation steps.
- 10 18. An apparatus as claimed in Claim 17, wherein the first mass analyzer (Q1) comprises a quadrupole mass analyzer.
19. An apparatus as claimed in Claim 17 or 18, wherein the final mass analyzer (Q3) comprises a quadrupole mass analyzer, and the first mass
15 analyzer (Q1), the linear ion trap (Q2) and the final mass analyzer (Q3) are axially aligned with one another.
20. An apparatus as claimed in Claim 17 or 18, wherein the final mass analyzer (Q3) comprises a time of flight device (40).
20
21. An apparatus as claimed in Claim 19 or 20, wherein the linear ion trap (Q2) includes a multipole rod set.
22. An apparatus as claimed in Claim 21, wherein the linear ion
25 trap (Q2) comprises a quadrupole rod set and wherein the rods of the mass analyzers (Q1 and Q3) and of the linear ion trap (Q2) have substantially similar radii and substantially similar spacings.
23. An apparatus as claimed in Claim 17, wherein each of the first
30 analyzer (Q1) and the final analyzer (Q3) comprise one of: a linear quadrupole; a linear time of flight analyzer (40); a reflectron time of flight analyzer; a single magnetic sector analyzer; a double focusing two sector mass analyzer having an electric sector and a magnetic sector; a Paul trap; a Wien filter; a Mattauch-Herzog spectrograph; an ion cyclotron mass spectrometer; and a Thomson

24. An apparatus as claimed in Claim 23, wherein the linear ion trap (Q2) includes a multipole rod set.

5

25. An apparatus as claimed in Claim 17 or 22, wherein the linear ion trap (Q2) has a pair of opposed x rods and a pair of opposed y rods, wherein a main RF drive (86) is connected to the x and y rods (88 and 90) of the linear ion trap (Q2), and wherein the auxiliary drive (84) is connected to at least one pair of rods of the linear ion trap (Q2).

26. An apparatus as claimed in Claim 25, wherein the auxiliary drive (84) is connected to the y rods (90) of the linear ion trap (Q2) through a transformer (85), and wherein the main RF drive (86) is connected directly to the x rods (88) of the linear ion trap (Q2) and, through a coil of the transformer (85) to the y rods (90).

27. An apparatus as claimed in Claim 25, which includes an
arbitrary waveform generator (82) connected to the auxiliary drive (84), for
20 applying a selected waveform to the linear ion trap (Q2) to excite ions therein.

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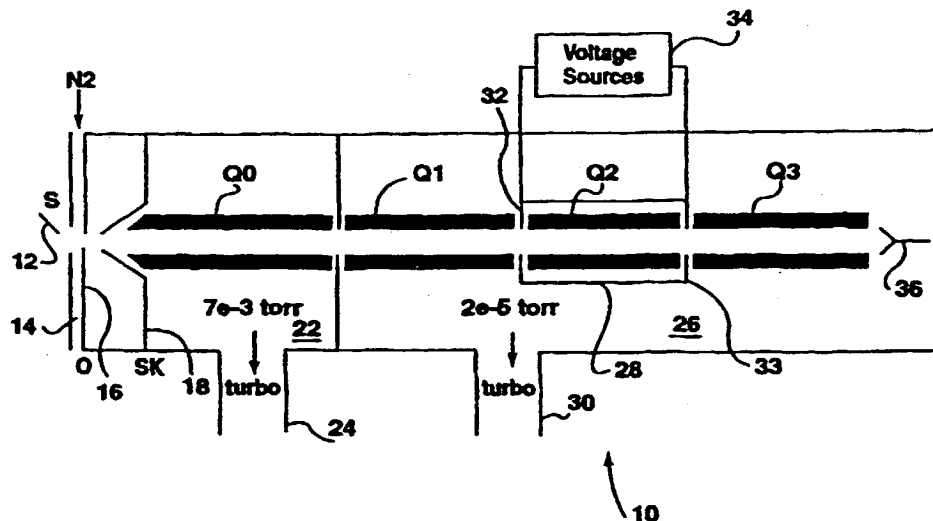
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(54) Title: METHOD AND APPARATUS FOR MULTIPLE STAGES OF MASS SPECTROMETRY



(57) Abstract

A method of and apparatus for analyzing a stream of ions first subjects a stream of ions to a first mass analysis step, to select ions having a mass-to-charge ratio in a first desired range; this enables a mass analyzer with high resolution to be used. The selected ions are then passed into a radiofrequency linear ion trap containing a gas. The trapped ions are caused to collide with the gas, either by being injected with a high axial energy or by application of external excitation to cause fragmentation. Fragment ions of a given mass-to-charge ratio can then be isolated and excited to produce fragments of fragments. This process can be repeated to give multiple steps of mass spectrometry, MSⁿ. The fragment ions, and undissociated precursors are then passed out of the linear ion trap and subjected to a further mass analysis step, for example in a time of flight device, to determine the mass spectrum of the ions.

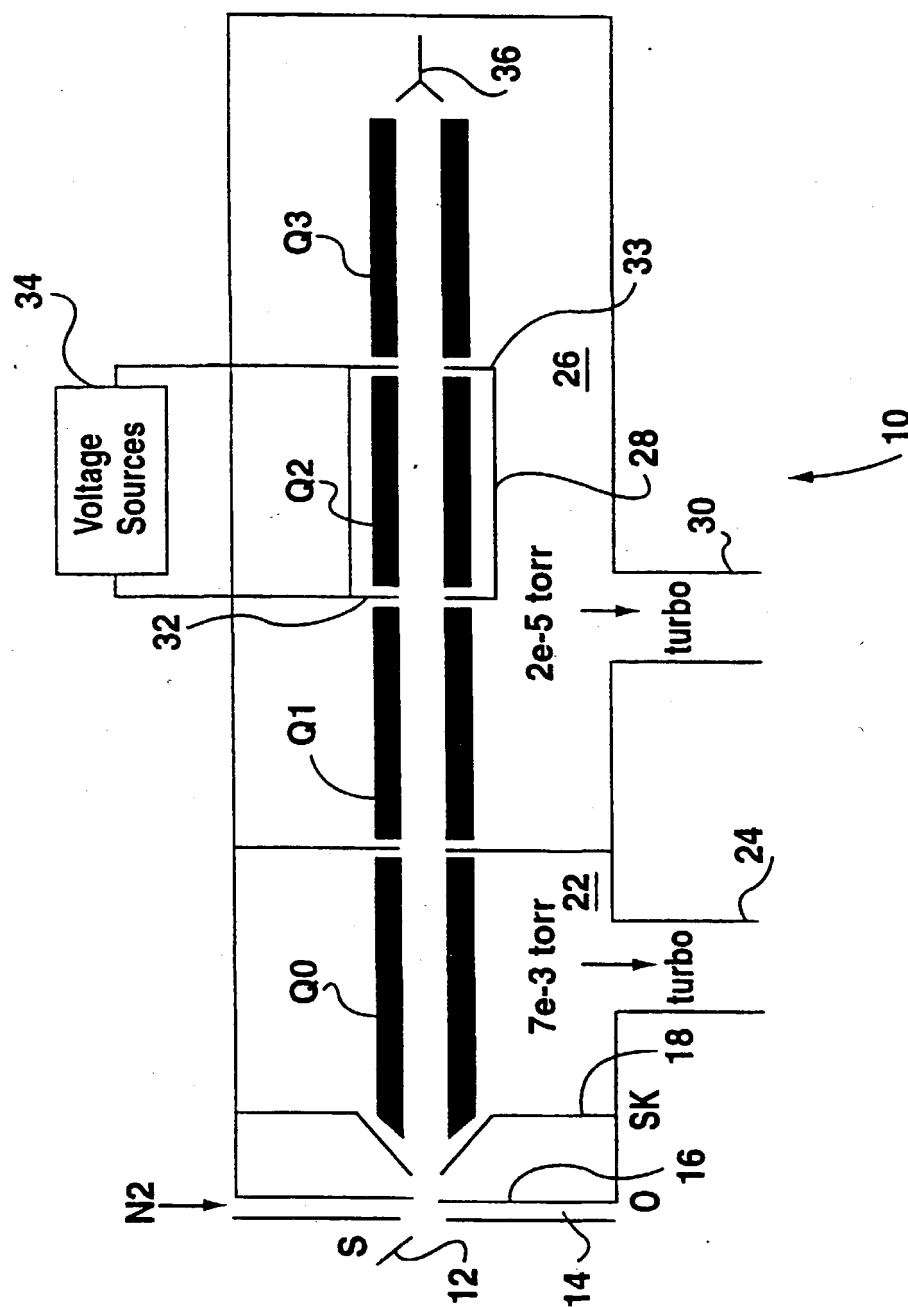


FIG. 1



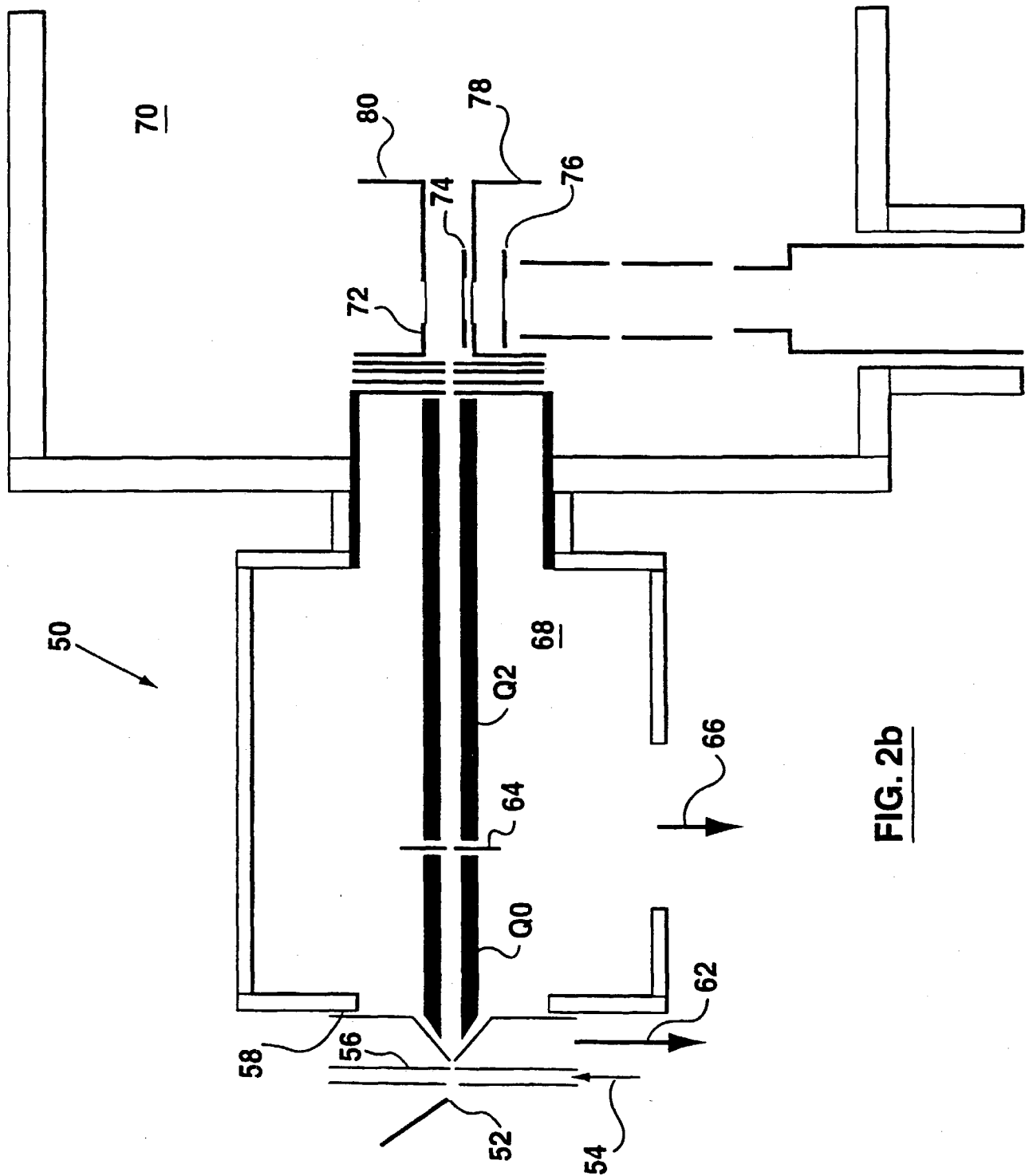


FIG. 2b

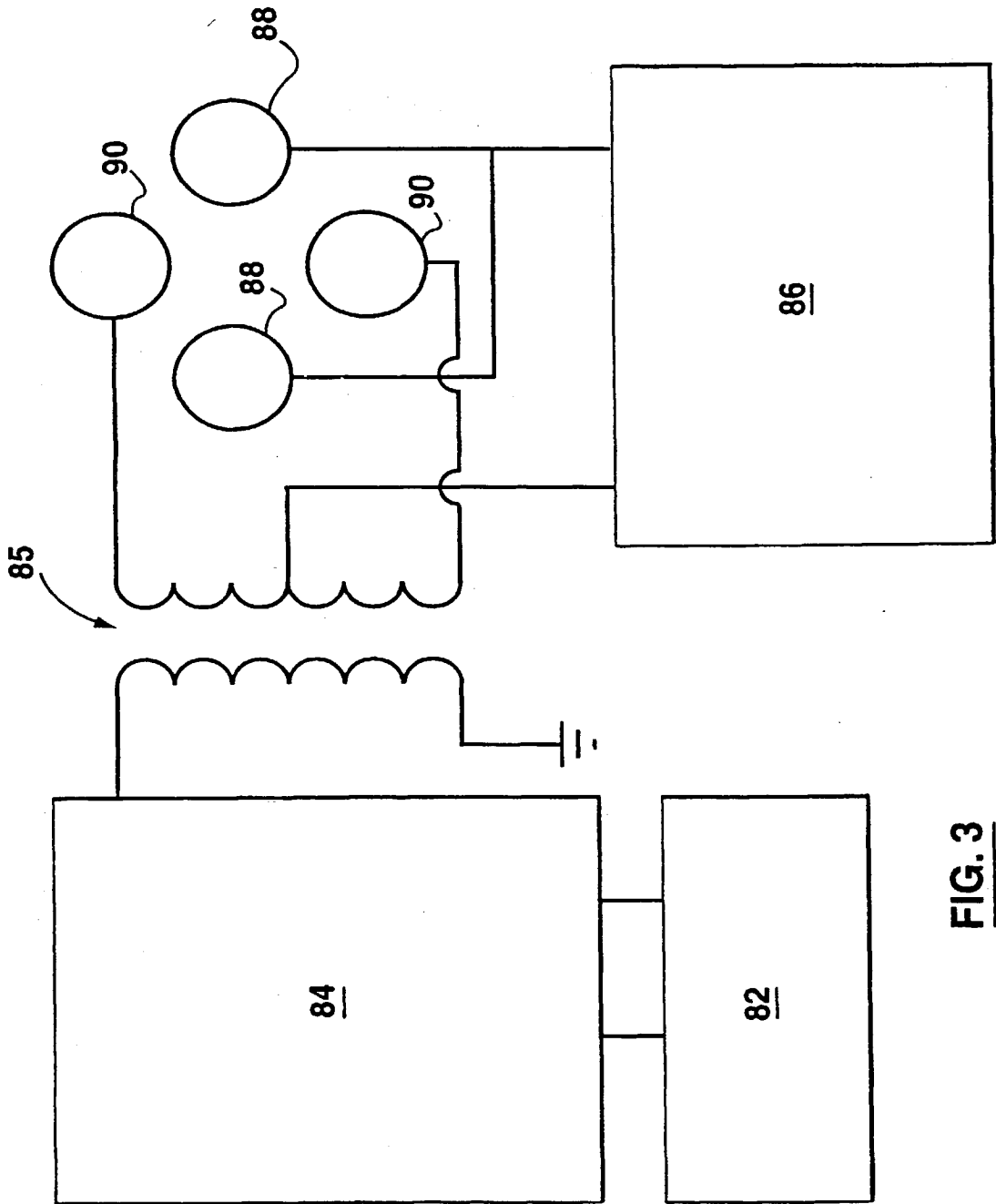


FIG. 3

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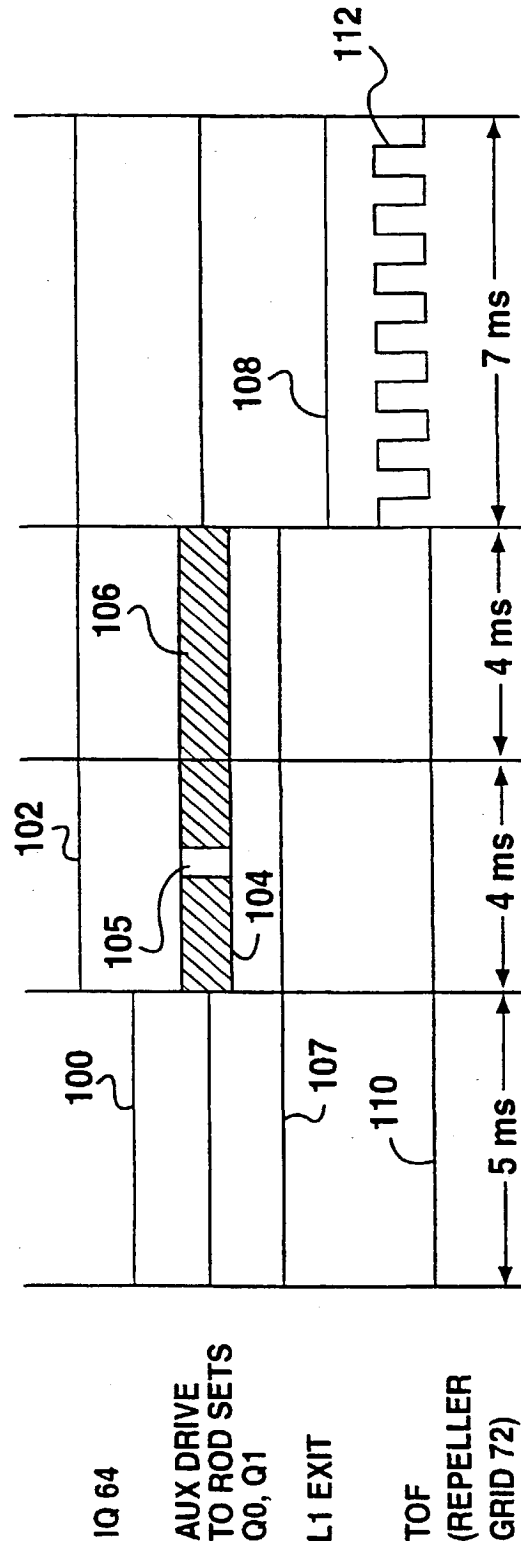
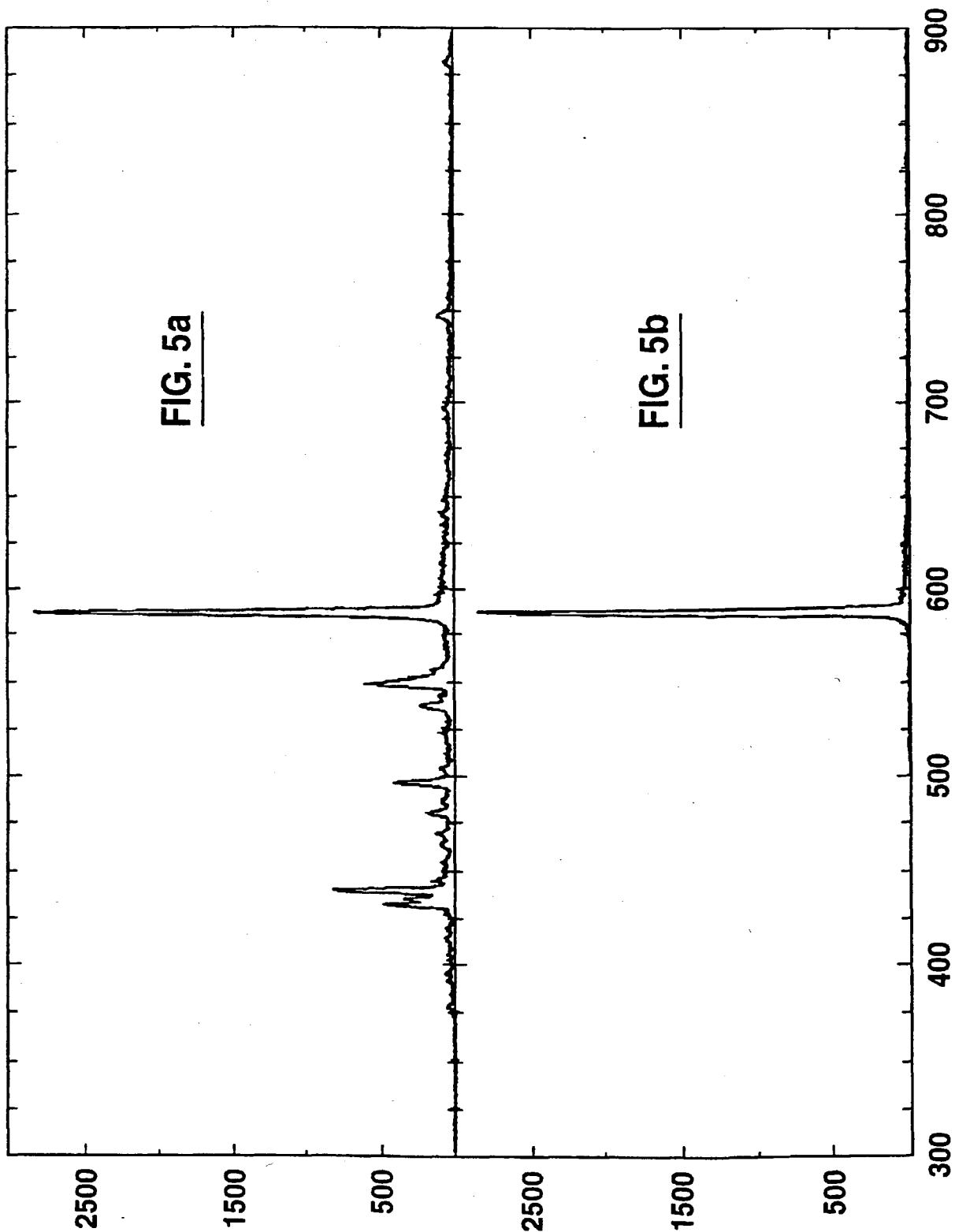
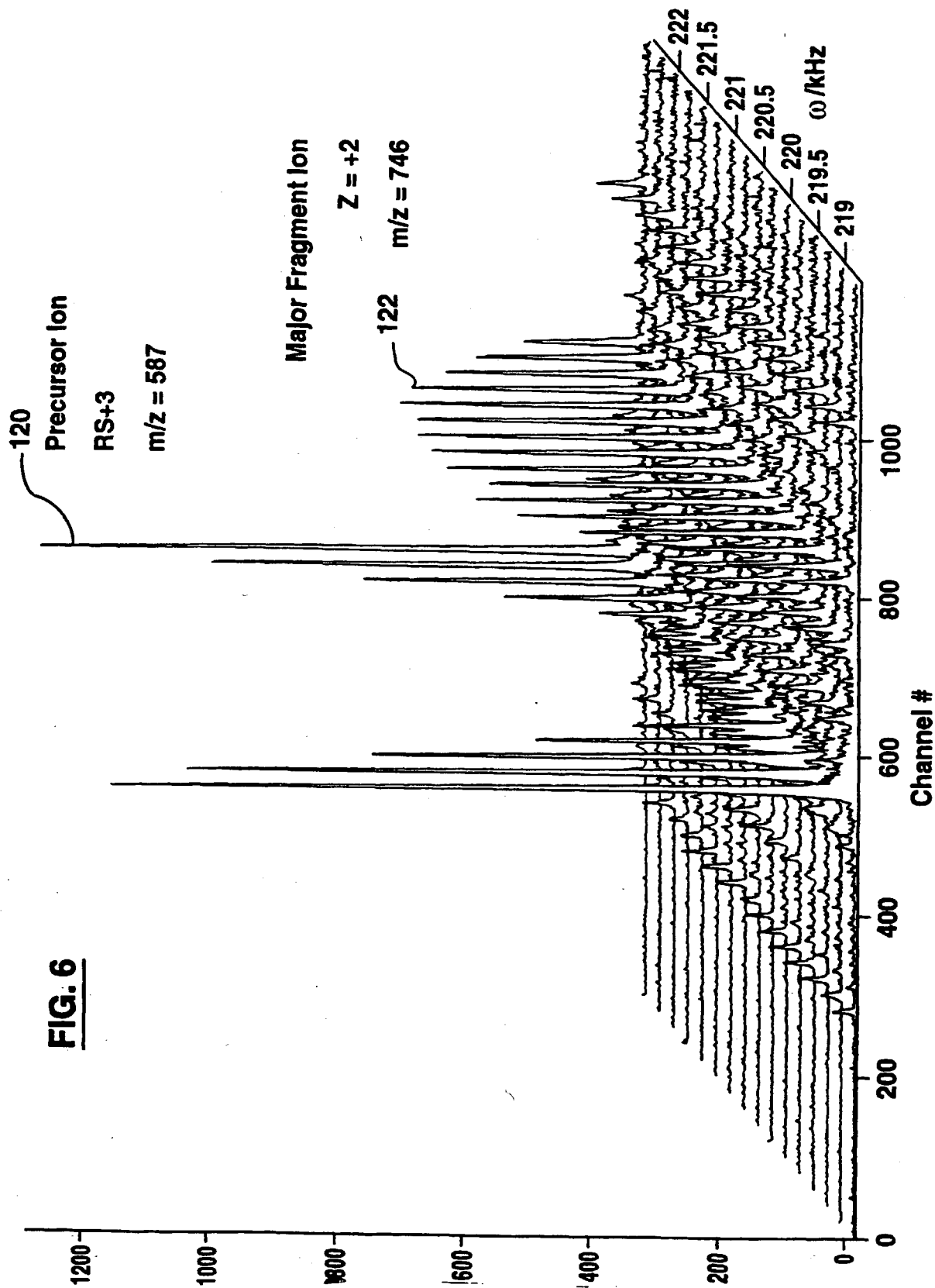


FIG. 4

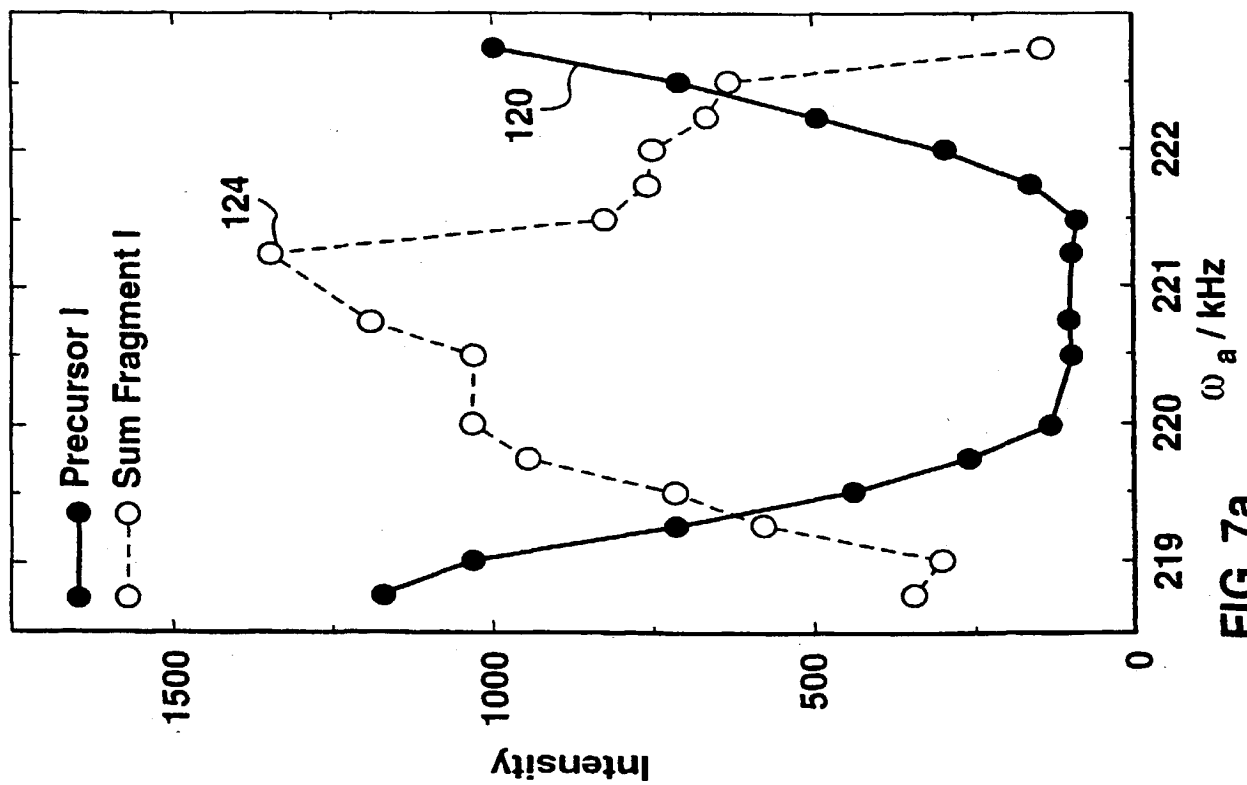
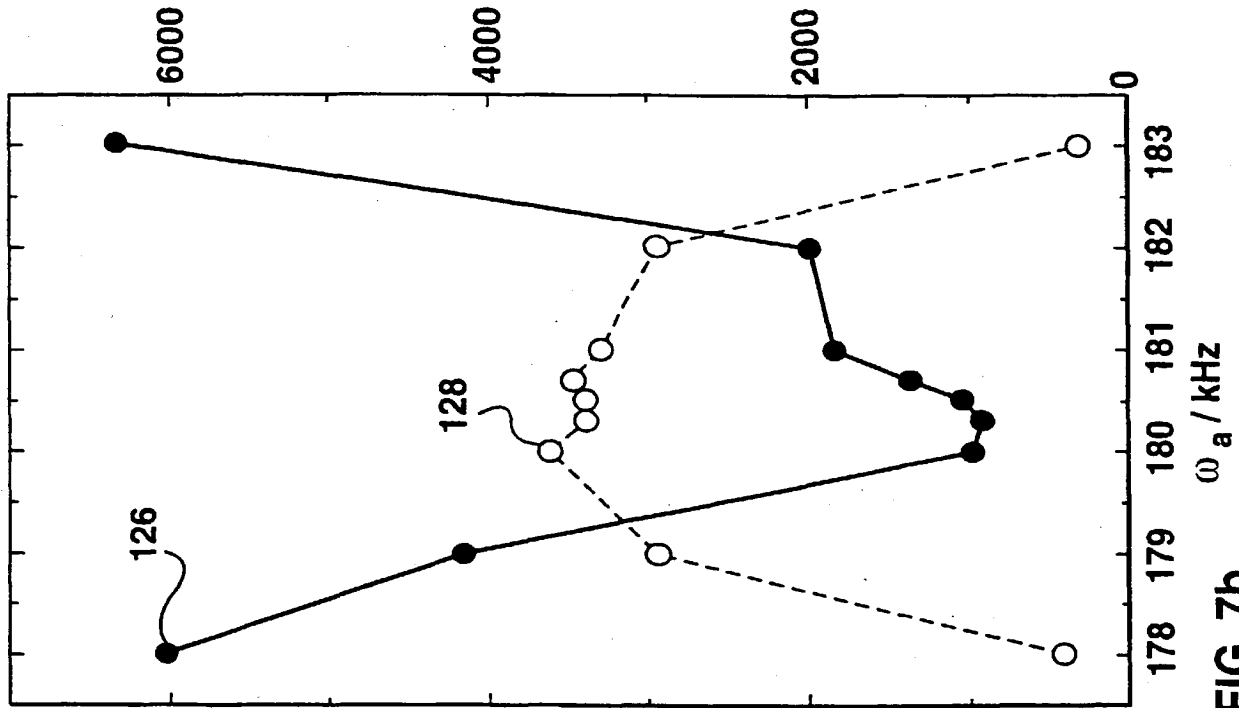
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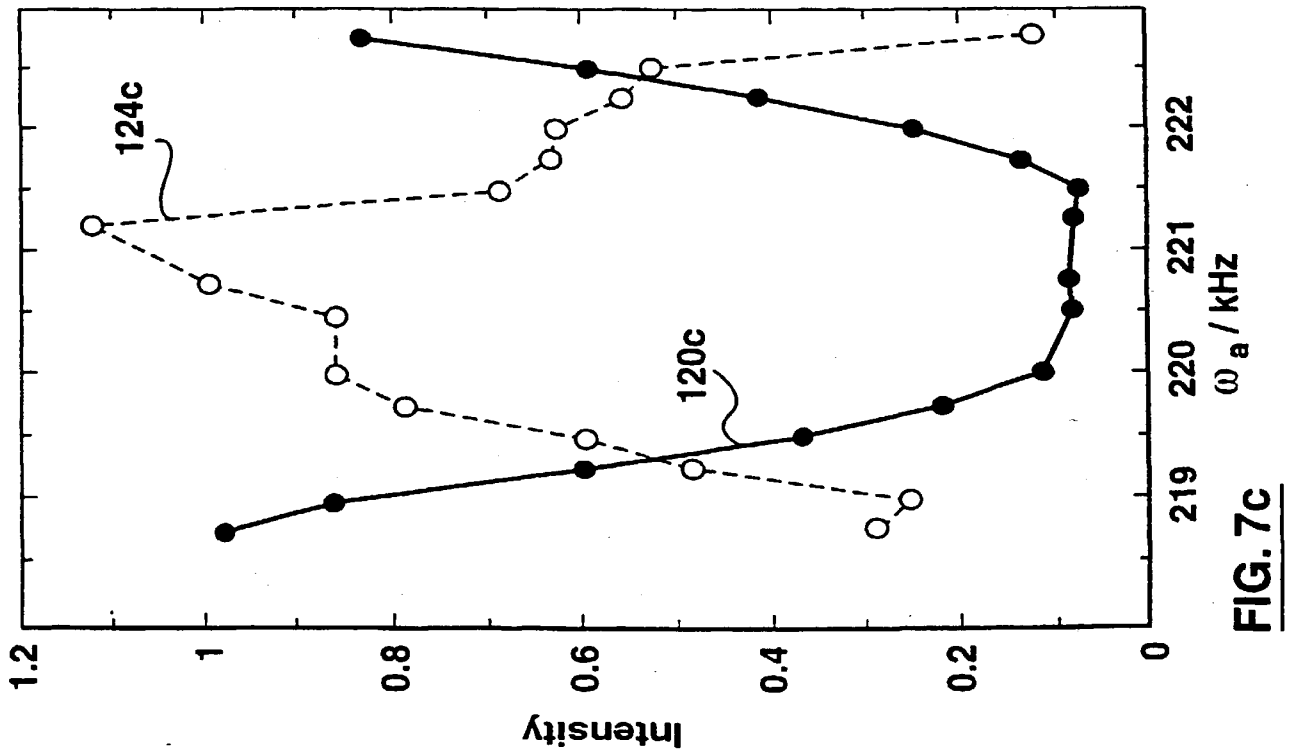
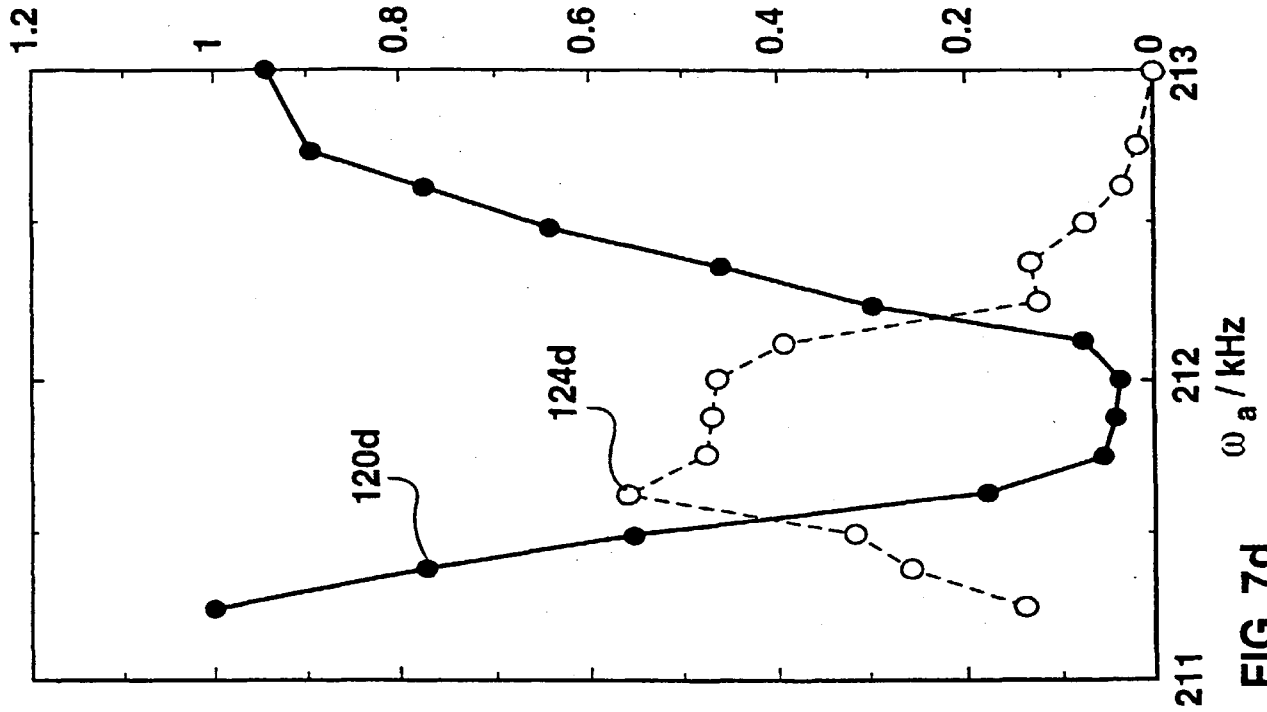
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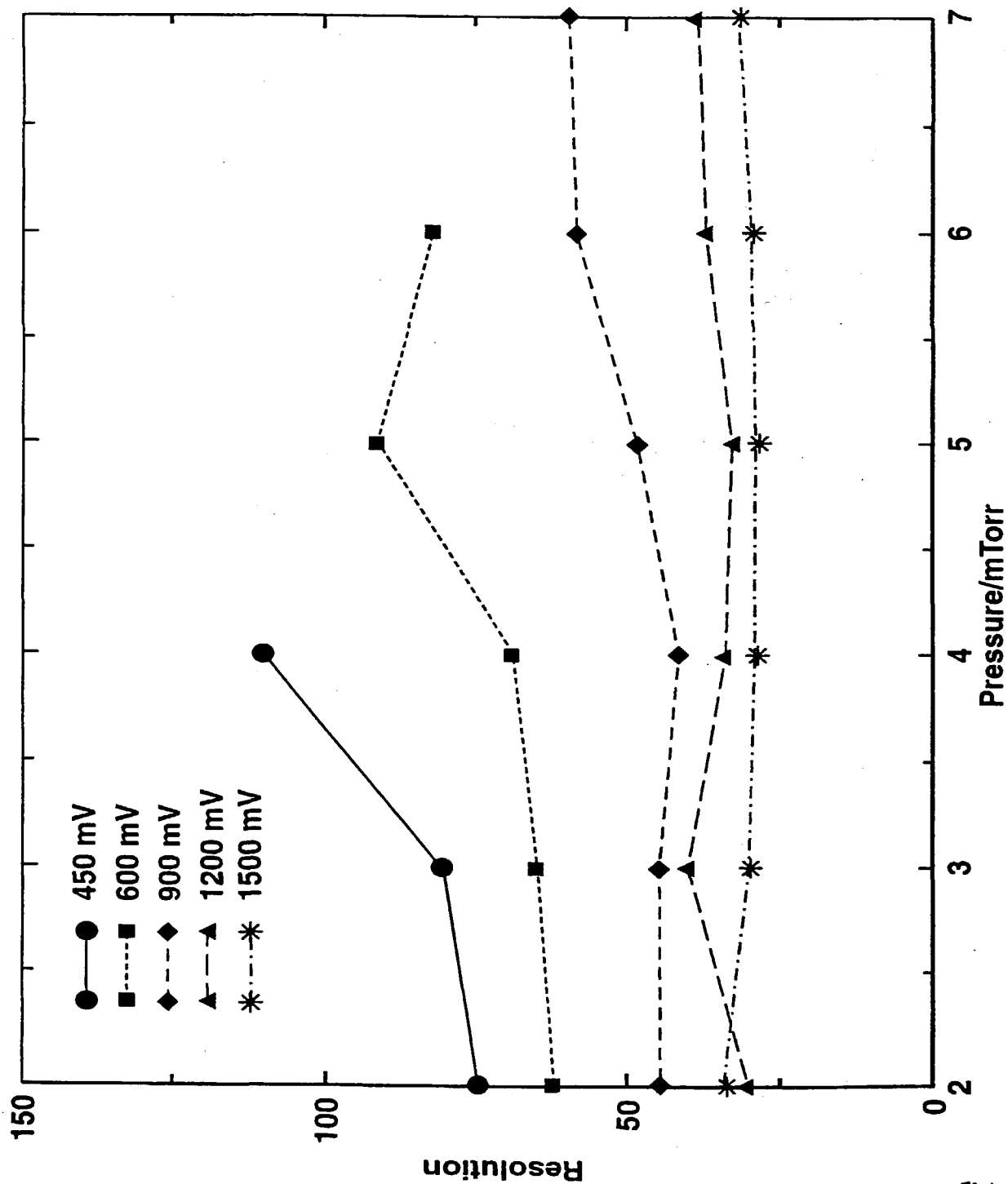
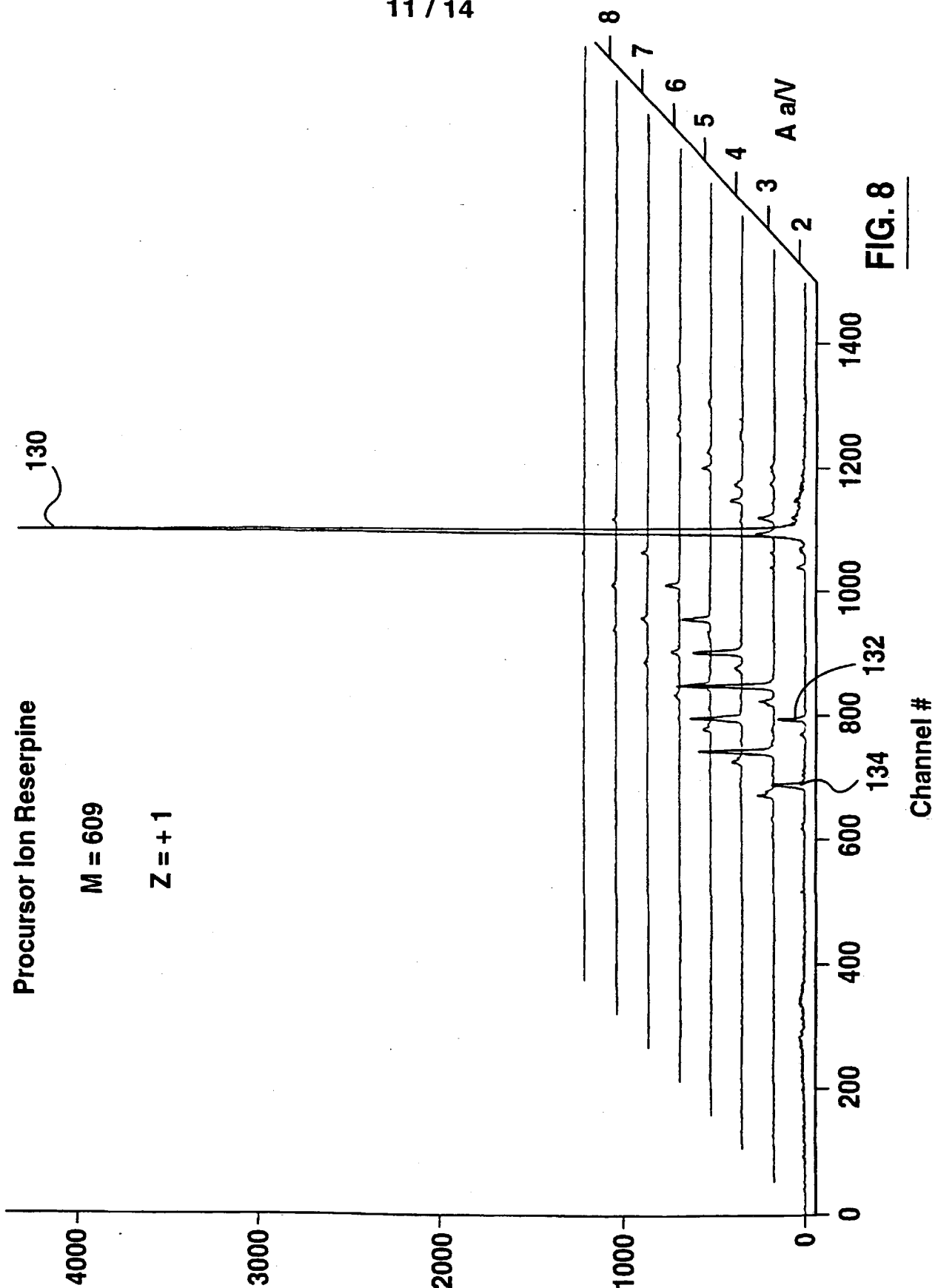
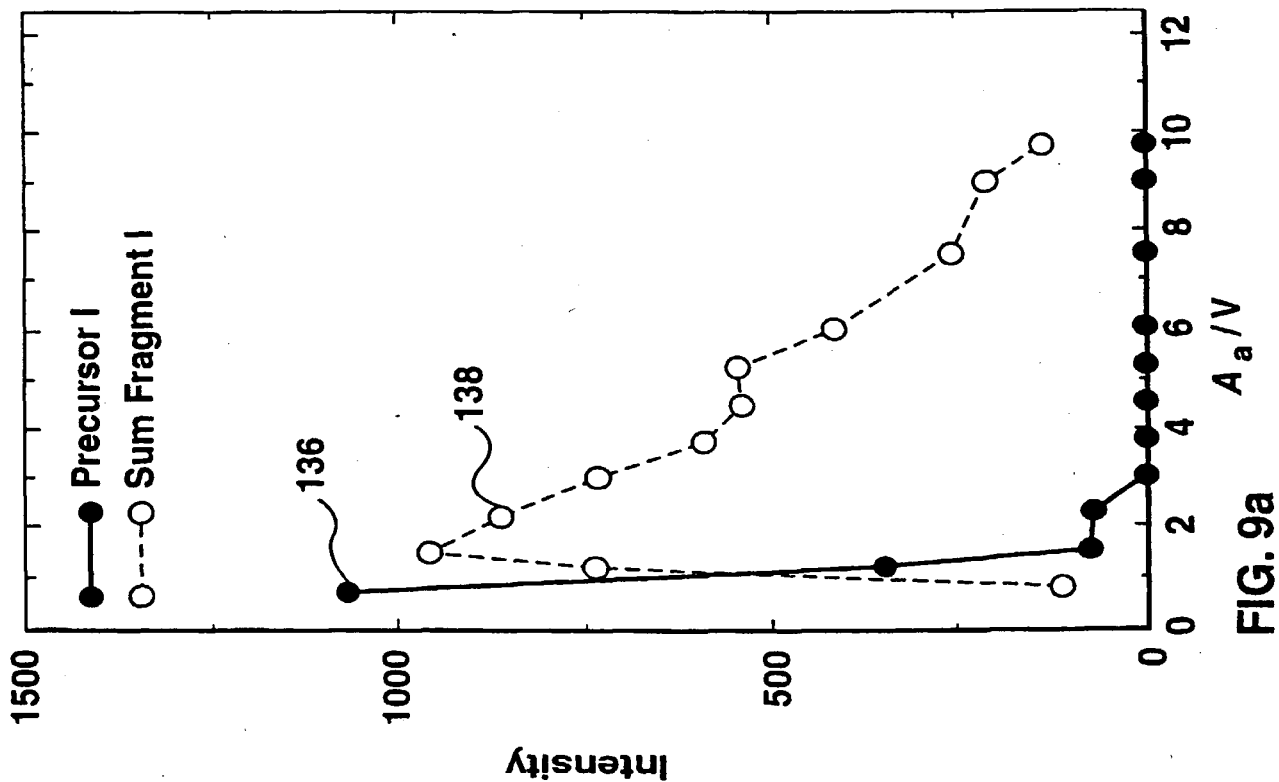
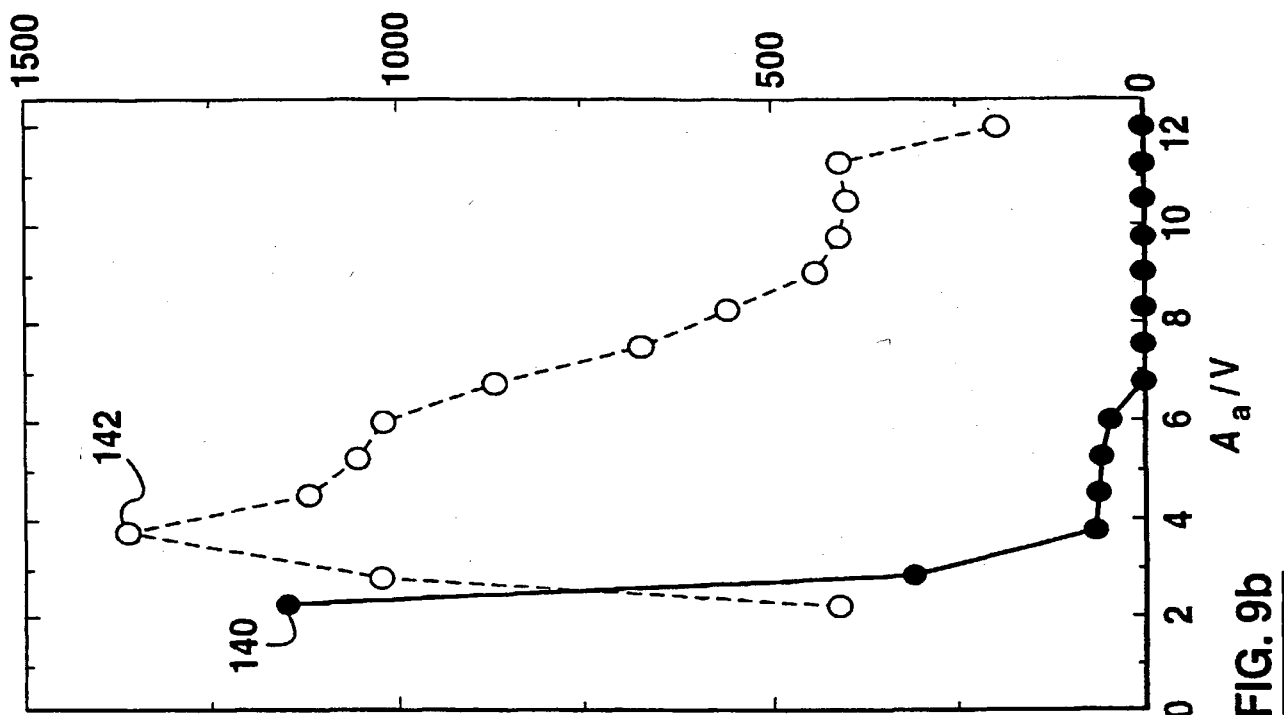


FIG. 7e

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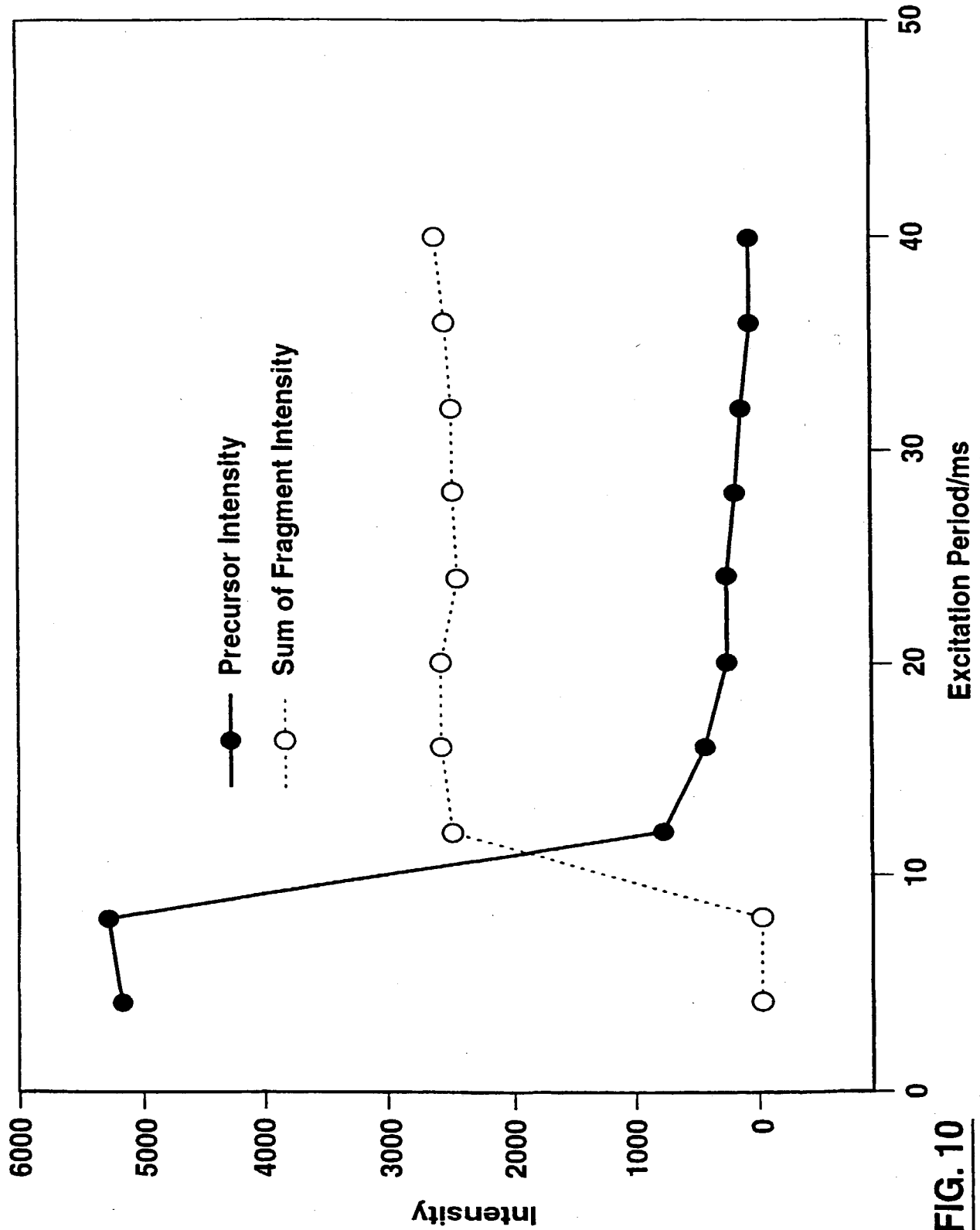
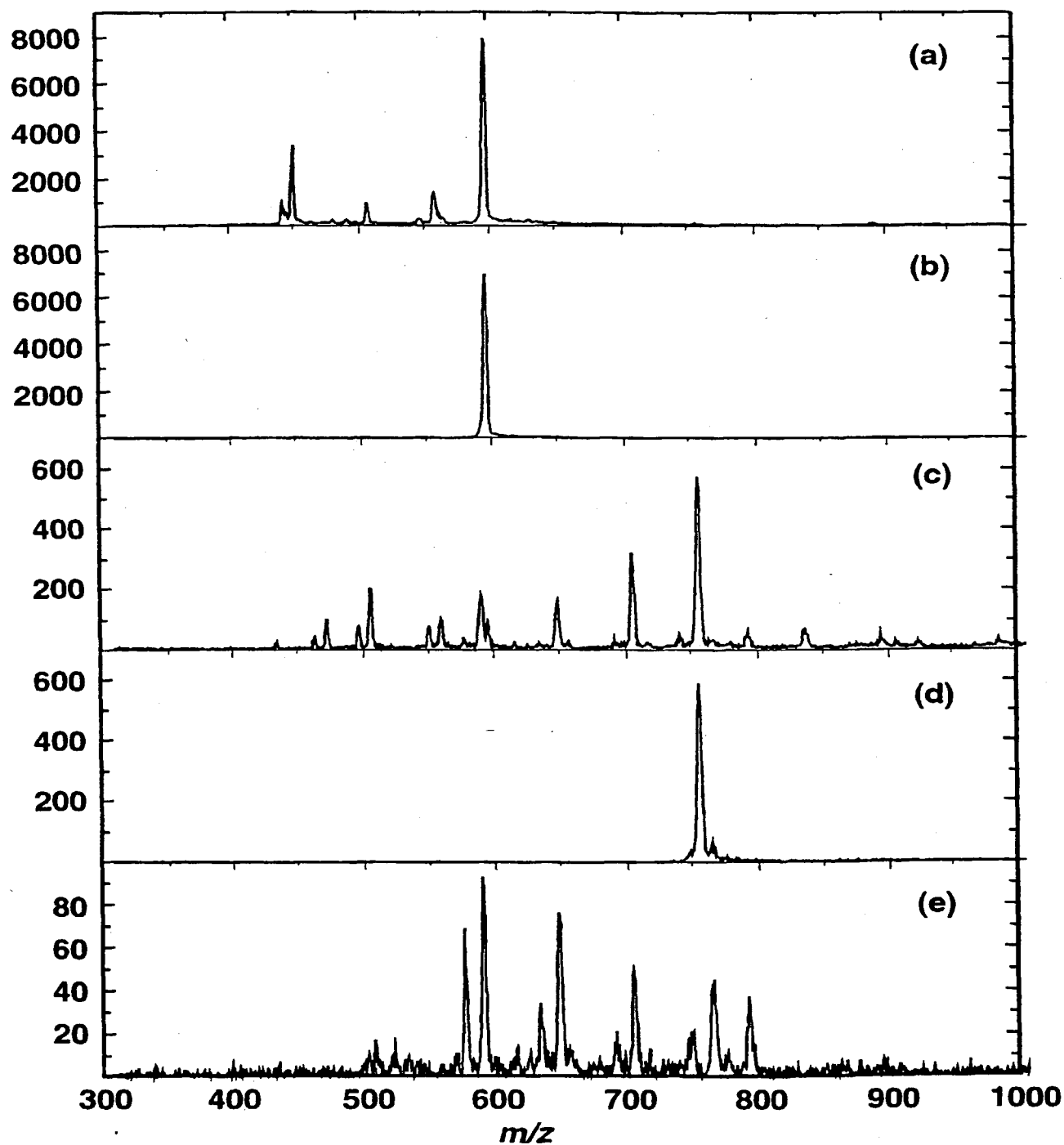


FIG. 10

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**FIG. 11**

PTO/SB/D1A (10-00)

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**DECLARATION (37 CFR 1.63) FOR UTILITY OR DESIGN APPLICATION USING AN
APPLICATION DATA SHEET (37 CFR 1.76)**

As the below named inventor(s), I/we declare that

This declaration is directed to:

- ☐ The attached application, or
☒ Application No 09/857,234 filed on June 1, 2001
☐ as amended on _____ (if applicable);

I/we believe that I/we am/are the original and first inventor(s) of the subject matter which is claimed and for which a patent is sought;

I/we have reviewed and understand the contents of the above-identified application, including the claims, as amended by any amendment specifically referred to above.

I/we acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me/us to be material to patentability as defined in 37 CFR 1.56, including material information which became available between the filing date of the prior application and the National or PCT International filing date of the continuation-in-part application, if applicable; and

All statements made herein of my/own knowledge are true, all statements made herein on information and belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like are punishable by fine or imprisonment, or both, under 18 U.S.C. 1001, and may jeopardize the validity of the application or any patent issuing thereon.

FULL NAME OF INVENTOR(S)Inventor one: Jennifer CampbellSignature: [Signature] Citizen of: CanadaInventor two: Bruce CollingsSignature: _____ Citizen of: CanadaInventor three: Donald J. DouglasSignature: _____ Citizen of: Canada

Inventor four: _____

Signature: _____ Citizen of: _____

☐ Additional inventors are being named on _____ additional form(s) attached hereto.

Burden Hour Statement: This collection of information is required by 35 U.S.C. 115 and 37 CFR 1.63. The information is used by the public to file (and the PTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This form is estimated to take 1 minute to complete. This time will vary depending upon the needs of the individual case. Any comments on the amount of time you are required to complete this form should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, Washington, DC 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Assistant Commissioner for Patents, Washington, DC 20231.

DECLARATION (37 CFR 1.63) FOR UTILITY OR DESIGN APPLICATION USING AN APPLICATION DATA SHEET (37 CFR 1.76)

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- ☒ The attached application, or
☐ Application No. _____, filed on _____,
☐ as amended on _____ (if applicable);

I/we believe that I/we am/are the original and first inventor(s) of the subject matter which is claimed and for which a patent is sought;

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All statements made herein of my/own knowledge are true, all statements made herein on information and belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like are punishable by fine or imprisonment, or both, under 18 U.S.C. 1001, and may jeopardize the validity of the application or any patent issuing thereon.

FULL NAME OF INVENTOR(S)

Inventor one: Jennifer Campbell Date: _____

Signature: _____ Citizen of: Canada

Inventor two: Bruce Collings Date: May 24, 2001

Signature: Bruce Collings Citizen of: Canada

Inventor three: Donald J. Douglas Date: May 30, 2001

Signature: Donald Douglas Citizen of: Canada

Inventor four: _____

Signature: _____ Citizen of: _____

☐ Additional inventors are being named on _____ additional form(s) attached hereto.

Burden Hour Statement: This collection of information is required by 35 U.S.C. 115 and 37 CFR 1.63. The information is used by the public to file (and the PTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This form is estimated to take 1 minute to complete. This time will vary depending upon the needs of the individual case. Any comments on the amount of time you are required to complete this form should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, Washington, DC 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Assistant Commissioner for Patents, Washington, DC 20231.